

## ***Ludwigia stolonifera* (Guill. & Perr.) P.H. Raven, Insight into its Phenotypic Plasticity, Habitat Diversity and Associated Species**

**Ashraf T. Soliman<sup>#</sup>, Rim S. Hamdy and Azza B. Hamed**

*Department of Botany and Microbiology, Faculty of Science, Cairo University, Giza, Cairo, Egypt.*

**T**HIRTY-TWO populations of *L. stolonifera* were monitored resulted in detection of 7 morphotypes in 5 different habitats. Wide spectrum species included *Phragmites australis*, *Eichhornia crassipes* and *Cyperus alopecuroides*. Cosmopolitan, Paleotropical and Pantropical elements contributed about 56.7% of the total number of the associated species. Therophytes constituted 46.7% while hydrophytes and helophytes 26.7%, hemicryptophytes, chamaephytes, geophytes and epiphytes were moderately represented. The similarity values between each pair of the 7 morphotypes based on the 42 macro and micro morphological characters were carried out. The fresh water morphotypes were correlated with the organic matter and turbidity, while the terrestrial morphotype was affected by carbonates and ammonia while the brackish and saline morphotypes were affected by electric conductivity, salinity, NaCl contents. *L. stolonifera* exhibits high degree of habitat diversity in the leaf shape and size; in the flower from typical bisexual 5-merous flowers with 10 stamens, to bisexual 6-merous flowers with 12 stamens; in size, position of bracteoles from basal to near base or near base and near or above; in the seed productivity/fruit from 60-80 seeds/fruit to 35-45 seeds/fruit from basal to near base or above; in the abundance of vesicles from abundant (up to 10 pneumatophores/cluster) to occasional vesicles (up to 3 pneumatophores/cluster); in the position of aperture from planaperturate to angulaperturate

**Keywords:** Flora, Hydrophytes, Clustering analysis, Morphological variation, Pneumatophore, Egypt.

### **Introduction**

The number of aquatic weeds in the Nile system of Egypt is about 35 species belong to 19 genera and 15 families (Täckholm, 1974 and Zahran & Willis, 1992). These plants are either entirely submerged, free-floating or their roots may penetrate the soil at the bottom stream. *Ludwigia stolonifera* is one of the most dominant species in watercourses in Egypt (Zahran & Willis, 1992).

Genus *Ludwigia* L. is represented in the Egyptian flora by two species, *L. stolonifera* (Guill. & Perr.) P.H. Raven and *L. erecta* (L.) Hara. The latter is very rare in Nile valley and canal banks (Täckholm, 1974). Some members of genus *Ludwigia* are highly invasive and cause significant economic and environmental damage in many parts of the world (Eppo, 2004 and Global Invasive Species Database, 2009). In Egypt, *Ludwigia stolonifera* tends to aggregate so dense to block the whole waterway, retarding the water flow, threaten plant biodiversity leading to habitat degradation. Accordingly, the study of its associated plants may reveal the diversity,

environmental behavior and current distribution of these plants. Floristic composition together with life form represented diagnostic characters in distinguish species especially hydrophytes which have wide geographical and ecological distribution.

The ability of an organism to change its phenotype in response to changes in the environment widely referred to phenotypic plasticity. Price et al. (2003) represented a fundamental way in which organisms cope with environmental variation, phenotypic plasticity encompasses all types of environmentally induced changes (e.g. morphological, physiological, behavioral and phenological) that may or may not be permanent throughout an individual's lifespan. Many aquatics occur in habitats characterized by strong environmental gradients so they often display high levels of polymorphism and phenotypic plasticity (Lacoull & Freedman, 2006) and display responses in their vegetative traits (Dorken & Barrett, 2004).

Recently, Amer et al. (2016) studied the macro- and micro-taxonomic characters of *L. stolonifera*, the study revealed the presence of seven

<sup>#</sup>Corresponding author email: ashsoliman@sci.cu.edu.eg

DOI: 10.21608/ejbo.2018.5268.1217

Edited by: Monier M. Abd El-Ghani, Faculty of Science, Cairo University, Cairo, Egypt.

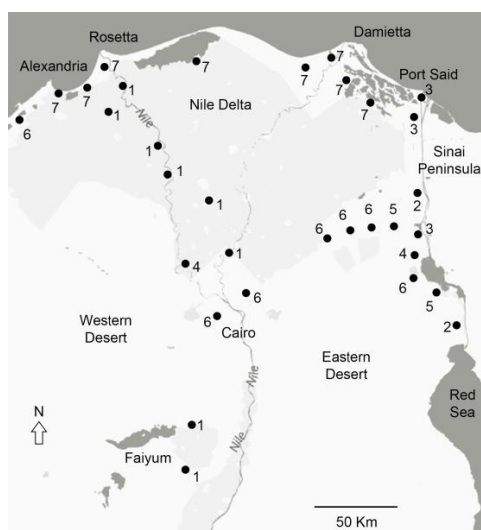
©2018 National Information and Documentation Center (NIDOC)

morphotypes. The current work aims to study the floristic composition associated with those seven morphotypes, the affinities and dissimilarities between each pair of those morphotypes, the phenotypic plasticity of *L. stolonifera* and factors determining this plasticity.

### Materials and Methods

An extensive survey was carried out between May 2013 and July 2015 at many localities covering different habitats in Egypt for collecting the available morphotypes of *Ludwigia stolonifera* and the associated plant species. These habitats included fresh, brackish, saline and salt-water habitats using a stratified sampling method (Müller-Dombois & Ellenberg, 1974).

Thirty-two populations (stands) of *Ludwigia stolonifera* were monitored (Fig. 1). These stands are distributed in 3 phytogeographical territories: 12 stands in the Mediterranean coast, 10 stands in the Nile valley and 10 stands in the Eastern desert. Each stand is 100m<sup>2</sup> (4x25m). Ecological notes, presence or absence of plant species had been recorded. The presence performance of a species was calculated as a percentage of the total number of stands where species recorded divided by the total number of stands. The stands were distributed as follows: M1 & M7 (8 stands for each), M6 (7 stands), M3 (3 stands) and M2, M4, M5 (2 stands for each) (Fig. 1).



**Fig. 1. Distribution map of the 32 populations (stands) representing the seven studied morphotypes (1-7) of *L. stolonifera* in Egypt.**

Identification of the associated species was carried out at Cairo University Herbarium (CAI) by the authors. Taxonomic nomenclature was according to Täckholm (1974) and Boulos (1995, 1999, 2000, 2002, 2005 and 2009). Growth or life form of the collected species was according to Raunkiaer's life form system (Raunkiaer, 1937). Analysis of phytogeographical ranges was carried out using Zohary (1947 and 1962). Samples of fresh material of *Ludwigia stolonifera* were collected during flowering and fruiting seasons. From each population, a random sample (at least 10 specimens) of flowering and fruiting plants were examined. For each morphological character, 10-20 samples were monitored. Mean value and standard deviation of each morphological character were calculated. Voucher specimens were kept in Cairo University Herbarium (CAI).

For pollen analysis, pollen grains were extracted from anthers collected from fresh floral buds, mounted on aluminum stubs, gold coated and viewed in a Jeol 1200 Ex II SEM at 20KV. Size measurements were obtained from the average of 10 randomly selected mature grains when possible. Among the investigated pollen morphological characters: pollen shape, pollen size, polarity, aperture position, aperture dimension (LxW), colpus length, exine sculpture and viscin threads diameter. The terminology used in pollen description followed Punt et al. (2007).

### Physicochemical analysis of the environmental factors

For water analysis, three water samples were collected from each stand from the top down to 50cm below the water surface, and then integrated as one composite sample. Three replicates were analysed for each sample. For soil analysis, three soil samples were collected from each stand at three depths: 0-10, 10-25 and 25-50cm. These samples were spread over sheets of paper, air dried, thoroughly mixed together, forming one composite sample, passed through 2mm sieve to remove gravel and debris. Three replicates were analysed for each sample. Soil extract were prepared (W/V, 1gm soil: 5ml distilled water) to meet the requirement for different determination.

Sixteen environmental variables were chosen to trace their effect on the phenotypic plasticity of *Ludwigia*. APHA (1995) Standard Methods for the Examination of Water and Wastewater were used in the determination of pH values using Method 4500-

H<sup>+</sup>, the electrical conductivity (which expresses the total soluble salts in water and soil extract) using Method 2510 B, salinity using Method 2520 B, turbidity using Method 2130 B, carbonates and bicarbonates using Method 2320 B and organic matter using Method 5220 B. Determination of chlorides, sulfates, phosphates and nitrates were carried out by ASTM (2011) D4327-11 standard test method for anions in water by suppressed ion chromatography. Determination of sodium, potassium, ammonium, magnesium and calcium were carried out using ASTM (2009) D6919-09 standard test method for determination of dissolved alkali and alkaline earth cations and ammonium in water by ion chromatography.

#### Data analysis

The basic purposes of the multivariate analyses are summarizing large complex data and refining models for habitat structures (Ward et al., 1993). Floristic data matrix (7 morphotypes in the monitored 32 stands and the associated 60 species) was subjected to classification by two-way indicator species analysis (TWINSPAN) using the default settings of the computer program, Community Analysis Package (CAP) for windows version 1.2. An ordered two-way table that expresses succinctly the relationships of the stands and species within the data set was constructed (Hill, 1979 and Økland, 1990). The stands are ordered first by divisive hierarchical clustering, and then the species are clustered based on the classification of stands (Gauch & Whittaker, 1981).

For studying the similarity between each pair of the morphotypes based on the macro- and micro-morphological characters (data matrix of 42 characters and 7 morphotypes) and that based on the associated species (60 species and 7 morphotypes), the data matrices were subjected to Jaccard's Measure of the program SPSS (version 20 for Windows).

In this study, the default options of the computer program CANOCO software version 4.5 (Ter-Braak & Smilauer, 2002) was used to check the magnitude of change in species composition along the first ordination axis (i.e., gradient length in standard deviation units). The canonical correspondence analysis (CCA) was chosen to be the appropriate ordination method to perform direct gradient analysis (Ter-Braak & Prentice, 1988). The relationships between floristic gradients and the studied environmental variables can be indicated on

the ordination diagram produced by CCA biplot. All data variables are assessed for normality (stustatw version 5; Berk, 1994) prior to the CCA analysis and appropriate transformations are performed. Sixteen environmental variables were included in this study: pH, electric conductivity (EC), salinity, turbidity, bicarbonates (HCO<sub>3</sub><sup>-</sup>), carbonates (CO<sub>3</sub><sup>-2</sup>), organic matter (OM), sodium (Na<sup>+</sup>), ammonium (NH<sub>4</sub><sup>+</sup>), potassium (K<sup>+</sup>), magnesium (Mg<sup>+2</sup>), calcium (Ca<sup>+2</sup>), chloride (Cl<sup>-</sup>), nitrate (NO<sub>3</sub><sup>-</sup>), phosphate (PO<sub>4</sub><sup>-3</sup>) and sulfates (SO<sub>4</sub><sup>-2</sup>). A Monte Carlo permutation test (99 permutations) was used to test for significance of the eigenvalues of the first canonical axis. Interset correlations from the CCAs were therefore used to assess the importance of the environmental variables.

#### Results

##### Floristic composition of the associated species

The monitored 32 populations of *Ludwigia stolonifera* (Fig. 1), were categorised in 5 different habitats; four aquatic viz. fresh water (M1 & M2), fresh to brackish (M4 & M6), brackish water (M5), marine water (M7) and one terrestrial wetland habitat (M3). The total number of the monitored associated species were 60 vascular plants belonging to 30 families. These are 29 annuals, 27 perennials and 4 trees or shrubs (Table 1).

The species richest families were Poaceae that was represented by 10 species, followed by Chenopodiaceae (6 species), Polygonaceae (5 species), Amaranthaceae (4 species) and Cyperaceae (3 species); they accounted for 46.7% of the total collected flora. Seven families were represented by 2 species each and 18 monogeneric families. *Amaranthus*, *Chenopodium*, *Cyperus* and *Persicaria* were the most species richest genera that were represented by 3 species each.

The number of associated species differed from one morphotype to another, the highest number was 32 species that were associated with the terrestrial wetland morphotype M3, followed by the marine morphotype (22 species), where the least number was recorded in association with the brackish morphotype M5 (6 species) followed by 7 species that were recorded in the fresh to brackish morphotype M4. Also, the presence performance of the associated species differed, it ranged between 75% (recorded in 24 stands out of 32) to about 3% (recorded in one stand). It is to be noted that 44 species were confined to only one stand (i.e., associated with only one of the 7 morphotypes).

TABLE 1. The presence performance % of the associated species within the different monitored morphotypes (M1-M7).

Morphotype Number	Species/total number of stands	Chorotype	LF	Hb	M1	M2	M3	M4	M5	M6	M7	T	%
	<i>Phragmites australis</i> (Cav.) Trin. ex Steud.	PAL	HH	P	4	1	2	2	2	6	7	24	75
	<i>Eichhornia crassipes</i> (C. Mart.) Solms	COSM	Hy	P	6	1	0	2	2	5	7	23	72
	<i>Cyperus alopecuroides</i> Rottb.	PAN	HH	P	4	1	1	1	0	2	1	10	31
	<i>Panicaria salicifolia</i> (Brouss. ex Willd.) Assenov	PAL	Hy	P	5	1	0	1	0	1	2	10	31
	<i>Myriophyllum spicatum</i> L.	COSM	Hy	P	0	0	0	1	1	2	1	5	16
	<i>Cynanchum acutum</i> L.	MED+IR-TR+EU-SB	Ep	P	0	1	1	0	0	2	1	5	16
	<i>Ceratophyllum demersum</i> L.	COSM	Hy	P	1	1	0	0	0	2	1	5	16
	<i>Potamogeton nodosus</i> Poir.	MED+IR-TR	Hy	P	1	1	0	0	0	1	0	3	9
	<i>Lemna gibba</i> L.	COSM	Hy	P	2	0	0	0	1	0	6	9	28
	<i>Ranunculus sceleratus</i> L.	MED+IR-TR+EU-SB	HH	A	2	0	0	0	1	0	2	5	16
	<i>Convolvulus arvensis</i> L.	PAL	Ep	P	0	1	1	0	0	2	0	4	13
	<i>Chenopodium ambrosioides</i> L.	COSM	Ch	P	1	0	1	0	0	0	0	2	6
	<i>Nymphaea lotus</i> L.	PAL	Hy	P	1	0	0	0	0	0	1	2	6
	<i>Cyperus rotundus</i> L.	PAN	G	P	0	0	2	0	0	1	0	3	9
	<i>Typha domingensis</i> (Pers.) Poir. ex Steud.	PAN	HH	P	0	0	1	0	0	0	4	5	16
	<i>Juncus acutus</i> L.	MED+IR-TR+EU-SB	H	P	0	0	1	0	0	0	1	2	6
	<i>Hordeum murinum</i> L. subsp. <i>leporinum</i> (Link) Arcang.	MED+SA-AR+EU-SB	Th	A	0	0	0	0	0	0	1	1	3
	<i>Mesembryanthemum nodiflorum</i> L.	MED+SA-AR+EU-SB	Th	A	0	0	0	0	0	0	1	1	3
	<i>Pisita stratiotes</i> L.	PAN	Hy	P	0	0	0	0	0	0	1	1	3
	<i>Cakile maritime</i> Scop. subsp. <i>aegyptiaca</i>	MED+SA-AR	Th	A	0	0	0	0	0	0	1	1	3
	<i>Spergularia marina</i> (L.) Griseb.	MED+IR-TR+EU-SB	H	P	0	0	0	0	0	0	1	1	3
	<i>Atriplex semibaccata</i> R. Br.	AUST	H	P	0	0	0	0	0	0	1	1	3
	<i>Halocnemum strobilaceum</i> (Pall.) M. Bieb.	MED+IR-TR+SA-AR	Ch	TS	0	0	0	0	0	0	1	1	3
	<i>Ipomoea cairica</i> (L.) Sweet	SA-AR	Ep	P	0	0	0	0	0	0	1	1	3
	<i>Imperata cylindrica</i> (L.) Raesch.	MED+IR-TR+SA-AR	H	P	0	0	0	0	0	0	1	1	3
	<i>Polygonum bellardii</i> All.	MED+IR-TR+EU-SB	Th	A	0	0	0	0	0	0	1	1	3

TABLE 1. Cont.

Species/total number of stands	Chorotype	M1								LF	Hb	M1	M2	M3	M4	M5	M6	M7	T	%				
		8	2	3	2	2	7	8	32															
<i>Zaleya pentandra</i> (L.) C. Jeffrey	S-Z	A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3			
<i>Cyperus laevigatus</i> L.	PAL	P	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3		
<i>Alhagi graecorum</i> Boiss.	PAL	P	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3	
<i>Wolffella hyalina</i> (Delile) Monod	SA-AR+S-Z	P	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Pluchea dioscoridis</i> (L.) DC.	SA-AR+S-Z	Ph	TS	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Chenopodium album</i> L.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Chenopodium murale</i> L.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Eclipta prostrata</i> (L.) L.	PAN	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Cynodon dactylon</i> (L.) Pers.	PAN	G	P	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Rumex dentatus</i> L.	MED+IR-TR	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Conyza bonariensis</i> (L.) Cronquist	MED	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Echinochloa crusgalli</i> (L.) P. Beauv.	PAN	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Amaranthus hybridus</i> L.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Amaranthus viridis</i> L.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Bassia indica</i> (Wight) A. J. Scott	S-Z+IR-TR	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Malva parviflora</i> L.	MED	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Polygogon monspeliensis</i> (L.) Desf.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Sisymbrium irio</i> L.	MED+IR-TR+EU-SB	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Portulaca oleracea</i> L.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Digitaria sanguinalis</i> (L.) Scop.	PAL	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Setaria viridis</i> (L.) P. Beauv.	COSM	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Amaranthus graecizans</i> L.	MED+IR-TR	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Melilotus indicus</i> (L.) All.	PAL	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Ricinus communis</i> L.	PAN	Ch	TS	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Dactyloctenium aegyptium</i> (L.) Willd.	PAL	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Tamarix nilotica</i> (Ehrenb.) Bunge	SA-AR+S-Z	Ph	TS	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3
<i>Datura innoxia</i> Mill.	PAN	Th	A	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	3

TABLE 1. Cont.

Morphotype Number	M1							M2							M3							M4							M5							M6							M7							T																																									
	LF							Hb							M1							M2							M3							M4							M5							M6							M7							T																											
Species/total number of stands	Chorotype							LF							Hb							M1							M2							M3							M4							M5							M6							M7							T																				
<i>Solanum nigrum</i> L.	COSM							Th							A							0							0							1							0							0							0							0							1							3													
<i>Panicum coloratum</i> L.	S-Z							H							P							0							1							0							0							0							0							0							0							1							3						
<i>Alternanthera sessilis</i> (L.) DC.	PAN							HH							A							1							0							0							0							0							0							0							1							3													
<i>Glinus lotoides</i> L.	PAL							Th							A							1							0							0							0							0							0							0							1							3													
<i>Plantago major</i> L.	COSM							Th							A							1							0							0							0							0							0							0							1							3													
<i>Persicaria lanigera</i> (R. Br.) Soják	TRP							HH							P							1							0							0							0							0							0							0							1							3													
<i>Persicaria senegalensis</i> (Meisn.) Soják	S-Z							HH							P							1							0							0							0							0							0							0							1							3													
<b>Total number of species</b>																						<b>15</b>							<b>9</b>							<b>32</b>							<b>7</b>							<b>6</b>							<b>11</b>							<b>22</b>																											

T= Total number of populations. Chorotype: COSM= Cosmopolitan, PAL= Palaeotropical, PAN= Pan-tropical, EU-SB= Euro-Siberian, IR-TR= Irano-Turanian, MED= Mediterranean, SA-AR=Saharo-Arabian, S-Z= Sudano-Zambeian, TRP= Tropical, AUST= Australia, LF= Life Form: Ep= Epiphytes, G= Geophytes, HH= Helophytes & Hydrophytes, Th= Therophytes, H= Hemicryptophyte, Ch= Chamaephyte, Hy= Hydrophyte, Ph= Phanerophyte, Hb= Plant Habit: A= Annual, P= Perennial, TS= Tree or shrub. Figures are the number of stands where the corresponding species is recorded.

Only the amphibious *Phragmites australis* was monitored to associate the 7 studied morphotypes, two species viz, the hydrophyte *Eichhornia crassipes* and the helophyte *Cyperus alopecuroides* were recorded in 6 of the studied morphotypes. The former was absent in the terrestrial wetland morphotype (M3), while the later was not recorded in the association of the brackish morphotype (M5). The distribution of the 44 confined species was as follows: 24 species (about 54.5% of the confined species) were confined to the terrestrial wetland morphotype (M3). Most of the latter were therophytes that can grow in the banks of canals, rivers and drains. 10 species in M7, 5 species in the fresh habitat of M1, two species in M4, one species (*Zaleya pentandra*) in M6, one species (*Cyperus laevigatus*) in M5, one species (*Panicum coloratum*) in M2.

Table 2 summarizes the chorological analysis of the associated species. Cosmopolitan, Paleotropical and Pan-tropical elements were represented by relatively high numbers 14, 10 and 10 species, respectively, that contributes about 56.7% of the total number of the associated species. Mediterranean element either mono, biregional and pluri-regional elements were represented by 2, 4 and 10 species, respectively that collectively represented 26.7%. Sudano-zambeian elements either mono or biregional were very poorly represented (3 and 4 species, respectively), while pure Saharo-Arabian element was also poorly represented by only one species.

The growth or life form of these associated species differed, The therophytes were relatively, highly represented by 28 species that constituted about 46.7% of the total number of associated species, followed by the hydrophytes and helophytes that were represented by 16 species about 26.7%. Hemicryptophytes were represented by only 5 species followed by chamaephytes, geophytes and epiphytes that were represented by 3 species each. *Tamarix nilotica* and *Pluchea dioscorides* were only the two phanerophytes recorded in the association of *Ludwigia stolonifera* in our study (Fig. 2).

The proximity matrix or the similarity between each pair of the 7 morphotypes based on their associated species (Table 3), scored its highest value in the combination between the two fresh to brackish morphotypes M4 & M6 (84%), and in the association between the marine (M7) and

brackish morphotype (M5). At the contrary, the least similarity values were recorded between the terrestrial wetland habitat M3 with the marine M7 (20%), the fresh water morphotype M1 (21%), the brackish M5 (21%) and the fresh to brackish M4 (23%) (Table 3).

The clustering analysis carried out by the CAP (community analysis Package), using agglomerative Ward's technique by Euclidean distance measure, on the macro and micro morphological matrix (42 characters) of the 7 studied morphotypes revealed that the 7 morphotypes were clearly distributed in 3 groups (Fig. 3). Group I included the fresh morphotypes M1 and M2, Group II included the brackish and marine the morphotypes M4-M7 and group III

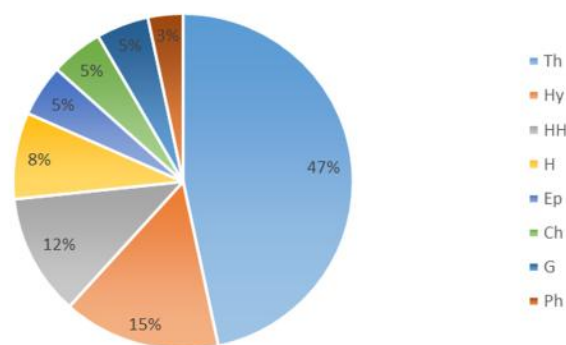
included the terrestrial morphotype M3.

In addition, Table 4 showed the similarity values between each pair of the 7 morphotypes based on the 42 macro and micro morphological characters. It was obvious that the highest similarity was recorded between the two fresh to brackish morphotypes M4 and M6 (66.7%) and between the two fresh morphotypes M1 and M2 (53.1%). At the contrary, terrestrial wetland morphotype recorded the least similarity values with the two fresh water morphotypes M1 and M2 (29.7% & 14.7%, respectively). The marine morphotype M7 recorded high similarity values; with other morphotypes ranged between 53.3% to 66.7% except for the terrestrial one M3 (32.4%).

**TABLE 2. Chorological analysis of the associated species.**

Chorotype	Number of species
Cosmopolitan	14
Palaeotropical	10
Pantropical	10
<b>Mediterranean</b>	
Monoregional	2
Biregional	4
Pleuroregional	10
<b>Sudano-Zambezian</b>	
Monoregional	3
Biregional	4
Tropical	1
Australian	1
Saharo-Arabian	1
<b>Total</b>	<b>60</b>

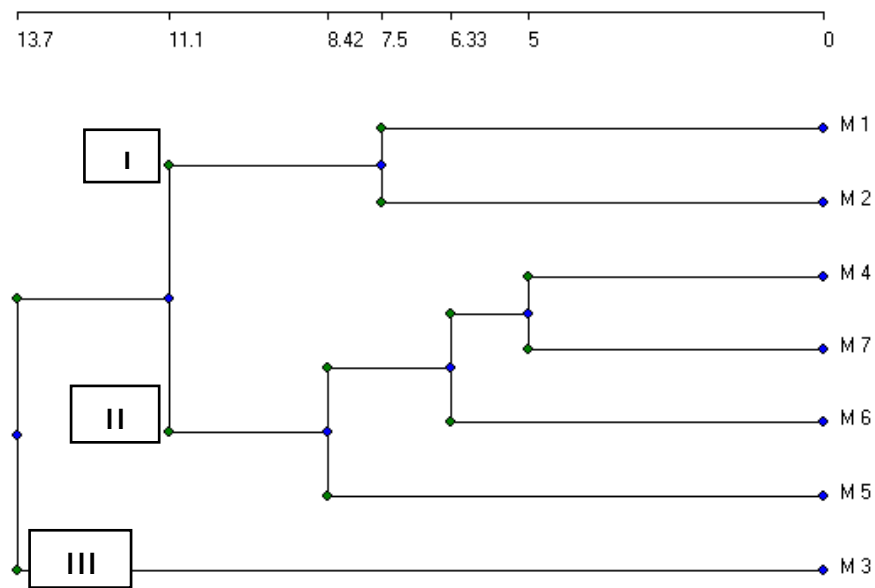
**Growth form of the associated species**



**Fig. 2. Different growth form of the recorded species (for abbreviations see Table 1).**

**TABLE 3. Proximity Matrix or the similarity between each pair of the 7 morphotypes (M1-M7) depending on their associated species.**

	M1	M2	M3	M4	M5	M6	M7
M1	100	74	<b>21</b>	80	73	80	78
M2		100	<b>26</b>	62	49	<b>81</b>	55
M3			100	<b>23</b>	<b>21</b>	40	<b>20</b>
M4				100	76	<b>84</b>	72
M5					100	80	<b>84</b>
M6						100	77
M7							100



**Fig. 3. Cluster analysis of the 7 morphotypes (M1-M7) by using agglomerative Ward's technique by Euclidean distance measure based on their macro and micro morphological data. I, II and III are the resultant groups.**

**TABLE 4. Similarity between morphotypes based on macro & micro morphological characters using Jaccard's Measure.**

	M1	M2	M3	M4	M5	M6	M7
M1	100						
M2	<b>53.1</b>	100					
M3	<b>29.7</b>	<b>14.7</b>	100				
M4	47.2	41.9	34.4	100			
M5	38.9	46.4	37.9	50	100		
M6	52.8	43.8	45.2	<b>66.7</b>	51.6	100	
M7	<b>61.8</b>	53.3	<b>32.4</b>	<b>66.7</b>	51.6	<b>57.6</b>	100



*Environmental variables-vegetation relationships*

Table 5 demonstrated the mean values, standard errors (SE) and ANOVA values (F-ratio & P=sig.) of the studied soil and water variables in each morphotype. Clearly, the water reaction (pH) of all samples is generally alkaline. Chlorides exhibited the most highly significant value (P=0.0007). In addition, electric conductivity, salinity, organic matter, potassium, magnesium, nitrates and sulfates were of high significant values. It also displayed that the stands of the saline morphotype M7 had the highest values of pH, EC, salinity,  $\text{HCO}_3^-$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{+2}$ ,  $\text{Ca}^{+2}$ ,  $\text{Cl}^-$ ,  $\text{NO}_3^-$  and  $\text{SO}_4^{-2}$ . Whereas, the organic matter and turbidity attained their highest values in the stands of the fresh morphotype M1,  $\text{NH}_4^+$  and  $\text{CO}_3^{-2}$  in stands of the terrestrial morphotype M3 and finally  $\text{PO}_4^{-3}$  in M4.

The application of CCA in the present data set demonstrated that electric conductivity, salinity, organic matter, potassium, magnesium, nitrates and sulfates were among the most important factors that affect the spatial distribution of the associated species of each morphotype. The relationship between the associated flora and water-soil analyses values in the 32 studied stands of the 7 morphotypes using CCA analysis was performed in Fig. 4. CCA ordination biplot of *L. stolonifera* morphotypes (1-7), with arrows represented the examined water-soil variables. Preliminary analysis revealed high inflation factors for 3 water variables,  $\text{K}^+$ ,  $\text{Mg}^{+2}$  and  $\text{Ca}^{+2}$  that should be excluded from the analysis.

It can be noticed that the stands of the fresh water morphotypes M1 & M2 were correlated the organic matter and turbidity, while the terrestrial morphotype M3 was affected by carbonates and ammonia while the brackish and saline morphotypes (M4, M5, M6 and M7) were affected by electric conductivity, salinity, NaCl contents.

The successive decrease of eigenvalues of the four CCA axes (0.2888, 0.4805 and 0.1092 for axes 1, 2 and 3, respectively) illustrated in Table 6, suggesting a well-structured data set. The species-environment correlations were high for the four axes, explaining 79.7% of the cumulative variance.

The inter-set correlations resulted from CCA of the examined water variables were shown in Table 6. Axis 1 was positively correlated with

$\text{CO}_3^{-2}$ ,  $\text{NH}_4^+$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{-2}$ , pH and negatively correlated with EC, Salinity, turbidity,  $\text{HCO}_3^-$ , organic matter,  $\text{Na}^+$ ,  $\text{Cl}^-$  and  $\text{PO}_4^{-3}$ . Carbonates and ammonia had the highest positive values (0.9951), while turbidity had the highest negative value (-0.7637), thus axis 1 can be interpreted as carbonates & ammonia-turbidity gradient. Axis 2 was positively correlated with pH, EC, Salinity,  $\text{HCO}_3^-$ ,  $\text{Na}^+$ ,  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{PO}_4^{-3}$ ,  $\text{SO}_4^{-2}$  and negatively correlated with Turbidity,  $\text{CO}_3^{-2}$ , OM and  $\text{NH}_4^+$ . The highest positive value was for  $\text{PO}_4^{-3}$  (0.6942), while the highest negative value was for organic matter (-0.7112), thus axis 2 can be interpreted as phosphate-organic matter gradient. Axis 3 was positively correlated with pH, EC, Salinity,  $\text{HCO}_3^-$ ,  $\text{CO}_3^{-2}$ , organic matter,  $\text{Na}^+$ ,  $\text{NH}_4^+$ ,  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{-2}$  and negatively correlated with Turbidity, and  $\text{PO}_4^{-3}$  thus axis 3 can be interpreted as Chloride-phosphate gradient.

The CCA biplot showed that the terrestrial morphotype M3 was correlated with carbonates and ammonia. This type of soil prefers the co-association of therophytes as *Chenopodium album*, *Chenopodium murale*, *Rumex dentatus*, *Amaranthus hybridus*, *Bassia indica*, *Malva parviflora* and *Melilotus indicus*. At the contrary, the increase in the electric conductivity, salinity and NaCl contents may result in presence or even dominance of tolerant species at the expense of less tolerant species as in the sites of the brackish and saline morphotypes (M4, M5, M6 and M7) that prefer the co-association of halophytes as *Spergularia marina*, *Atriplex semibaccata*; hydrophytes as *Pistia stratiotes*, *Zaleya pentandra*, *Wolffiella hyalina* and canal bank species as *Imperata cylindrica*, *Polygonum bellardii*, *Cyperus laevigatus*.

A test for significance with an unrestricted Monte Carlo permutation test found the F-ratio for the eigenvalue of CCA axis 1 and the trace statistics to be significant (P=0.04), indicating that the observed patterns did not arise by chance.

*Phenotypic plasticity of L. stolonifera**Habitat diversity*

*Ludwigia stolonifera* exhibits high degree of habitat diversity, this plasticity enables it to expand its ecological range from fresh water habitats as in morphotypes M1 and M2, wetlands M3, to brackish water M5 as well as marine habitats M7.

TABLE 5. Mean value, standard error and ANOVA F-ratio &amp; Sig. (P) values of the soil &amp; water variables in the 7 morphotypes.

Var	M1	M2	M3	M4	M5	M6	M7	F	Sig.
PH	7.71±.08	6.29±.10	7.89±.02	7.85±.03	8.30±.03	8.06±.03	8.66±.04	1.410	0.2861
EC	463.33±.88	553.61±1.32	619.12±34.81	775.61±8.34	1227.12±1.15	702.16±1.2	5436.56±4066.41	14.310	0.0030**
Sal	.24±.01	.39±.02	.41±.22	.89±.34	5.65±2.42	1.3±.63	38.33±3.12	12.620	0.0034**
Tur	4.22±.06	1.71±.02	.00	1.01±.12	2.11±.070	2.38±.08	2.00±.08	3.190	0.0925
HCO <sub>3</sub>	203.78±2.04	183.03±2.03	69.85±1.74	299.17±32.24	137.44±4.31	113.65±5.04	352.25±4.11	.642	0.5218
CO <sub>3</sub>	.00	.00	86.31±2.61	.00	.00	.00	.00	.897	0.6945
OM	253.4±2.24	9.84±.10	21.06±.53	11.09±.58	19.12±.59	44.22±.40	134.04±.58	16.603	0.0026**
Na	41.06±1.16	95.27±1.47	145.82±.43	172.94±.58	297.48±.29	178.73±2.03	2457.42±9.81	5.212	0.0216*
NH <sub>4</sub>	.00	.00	41.05±3.46	.00	.00	.00	.00	1.498	0.2791
K	5.65±.33	7.48±.29	14.36±.32	17.55±.29	6.34±.33	5.53±.29	79.34±.33	16.739	0.0021**
Mg	12.61±.31	17.54±.29	49.61±.31	23.48±.29	15.62±.31	13.39±.31	294.55±.29	11.797	0.0038**
Ca	31.53±.29	45.52±.29	138.43±.30	55.43±.30	50.52±.29	36.93±.58	181.58±.30	2.2794	0.1623
Cl	25.56±.29	57.38±.31	360.36±.32	324.6±.31	655.42±.30	324.66±.33	4154.24±1.75	19.450	0.0007***
NO <sub>3</sub>	.15±.01	.13±.00	33.49±.29	25.57±.30	.24±.01	1.20±.02	6.55±.29	13.713	0.0032**
PO <sub>4</sub>	.00	.00	.00	.37±.03	.00	.00	.00	.195	0.2127
SO <sub>4</sub>	34.4±.29	185.83±.58	632.37±.88	216.16±.48	239.53±1.44	201.74±1.11	889.09±.58	15.126	0.0027**

\* = P ≤ 0.05, \*\* = P ≤ 0.01, \*\*\* = P ≤ 0.001, EC= Electric conductivity, OM=Organic matter (ppm), Sal = Salinity (ppt) and Tur = Turbidity (NTU). Other variables are measured in mg/L.

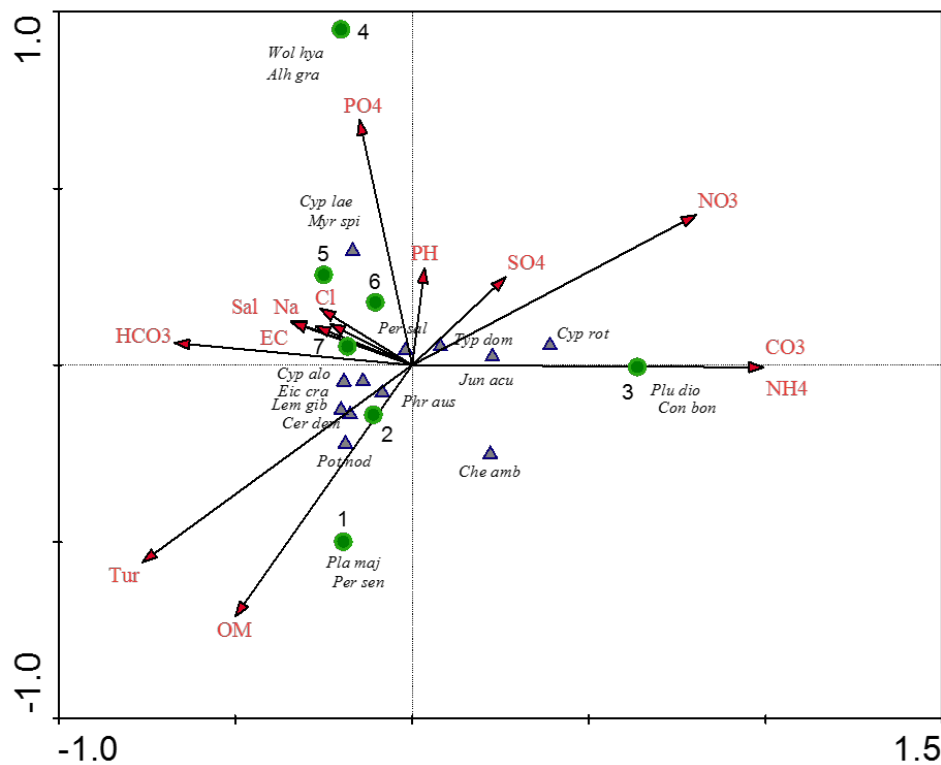


Fig. 4. The CCA ordination biplot of the first 2 axes showing the distribution of 7 morphotypes, with the examined environmental variables. Characteristic species are given with the 3 initial letters, for species names see Table 1.

TABLE 6. CCA analysis results showing the inter-set correlations of the water variables, together with eigenvalues and species-environment correlation of the studied morphotypes.

Axes	Ax1	Ax2	Ax3
<b>Eigenvalues</b>	0.2888	0.4805	0.1092
<b>Species-environment correlations</b>	44.5	64.5	79.7
pH	0.0359	0.2745	0.5199
EC	-0.3423	0.1245	0.9115
Salinity	-0.3419	0.1211	0.9123
Turbidity	<b>-0.7637</b>	-0.5572	-0.2049
HCO <sub>3</sub> <sup>-</sup>	-0.6722	0.0626	0.6264
CO <sub>3</sub> <sup>-2</sup>	<b>0.9951</b>	-0.0060	0.0309
OM	-0.4998	<b>-0.7112</b>	0.1414
Na <sup>+</sup>	-0.3145	0.1251	0.9215
NH <sub>4</sub> <sup>+</sup>	<b>0.9951</b>	-0.0060	0.0309
Cl <sup>-</sup>	-0.2738	0.1104	<b>0.9366</b>
NO <sub>3</sub> <sup>-</sup>	0.8045	0.4255	0.0095
PO <sub>4</sub> <sup>-3</sup>	-0.1492	<b>0.6942</b>	<b>-0.3448</b>
SO <sub>4</sub> <sup>-2</sup>	0.2651	0.2493	0.9205

#### Leaf diversity

Phenotypic plasticity reflected on the leaf shape and size of the studied morphotypes of *L. stolonifera* varied from narrowly elliptic-lanceolate in fresh water habitat as in M1, spatulate-narrowly elliptic in wetland habitat

M3 to lanceolate in marine habitat (M7). Moreover, diversity was clear in leaf sizes from relatively very small with mean value  $0.9 \pm 0.11$  cm in wetland morphotype M3 to relatively large with mean value  $11.3 \pm 1.7$  cm in fresh aquatic morphotype M1 (Fig. 5).



Fig. 5. Leaf diversity of different morphotypes (M1-M7) of *L. stolonifera*, each square =1cm.

#### Flower diversity

The studied morphotypes revealed notable variation in the flower from typical bisexual 5-merous flowers with 10 stamens as in M1 and M7, to bisexual 6-merous flowers with 12 stamens as in M3. Female flowers was traced also on some individuals with the bisexual flowers as in M2, M4, M5 and M6. In addition, the ovary shows variation in number of locules from typical 5-locules as in M1, M4, M6 and M7, to 5-8 locules in the other morphotypes (Fig. 6).

#### Fruit diversity

The fruit morphology of *L. stolonifera* showed notable variation in size, position of bracteoles and number of seeds/fruit among the different morphotypes from small fruit that may reach 2.5cm in M3 to relatively large fruit that may reach 3.5 in other morphotypes except M5. The position of bracteoles varied from basal in M1, M2 and M5 to near base in M4 and M6, at or near base in M3 and near or aboved in M7 (Fig. 7). Also, the seed productivity / fruit varies from 60-80 seeds/fruit in M1, M2 & M7 to 35-45 seeds/fruit in others.

#### Pneumatophores diversity

The studied morphotypes showed variation in the production of the pneumatophores (vesicles) at the nodes of floating stem, from abundant vesicles (up to 10 pneumatophores/cluster) as in M1, M2, M4 and M7 (Fig. 8.a & b) to occasional vesicles

(up to 3 pneumatophores/cluster, Fig. 8c) in the other morphotypes. Abundant Pneumatophores varied from conical to cylindrical (2.0-4.5(-9.0) cm) in M1, to conical to fusiform (2.0-4.5cm) in M2, fusiform (2.0-6.0cm) in M4 and conical to fusiform or cylindrical (2.0-4.5(-15.0)cm) in M7 where conical and cylindrical pneumatophores can be seen in the same cluster with branched pneumatophores on the roots (Fig. 8d). Occasional pneumatophores appeared fusiform in M5. The density of pneumatophores (number of pneumatophores at each node) differed. It was abundant (from 6 to 10) in M1, M2, M4 and M7 and occasional (up to 3) in others.

#### Pollen grains

The ratio of the pollen length in the saline morphotype M7,  $P/E = 0.89$  was relatively small compared with the fresh morphotypes  $P/E = 1.03$ . As well, M7 represented characteristic position of aperture, planaperturate while others were angulaperturate. Another example of plasticity is the colpus length that might reach  $31 \mu\text{m}$  in M4 and the aperture was covered by an operculum while in others (M2, M5 and M7), the colpus length might be less than  $7 \mu\text{m}$  and the aperture was without that operculum. In addition, the crystal-like elements on exine surface were observed in M3 and M5, and were absent in the other morphotypes. Moreover, family Onagraceae is characterized by the presence of viscin threads that were observed in all morphotypes except M5.

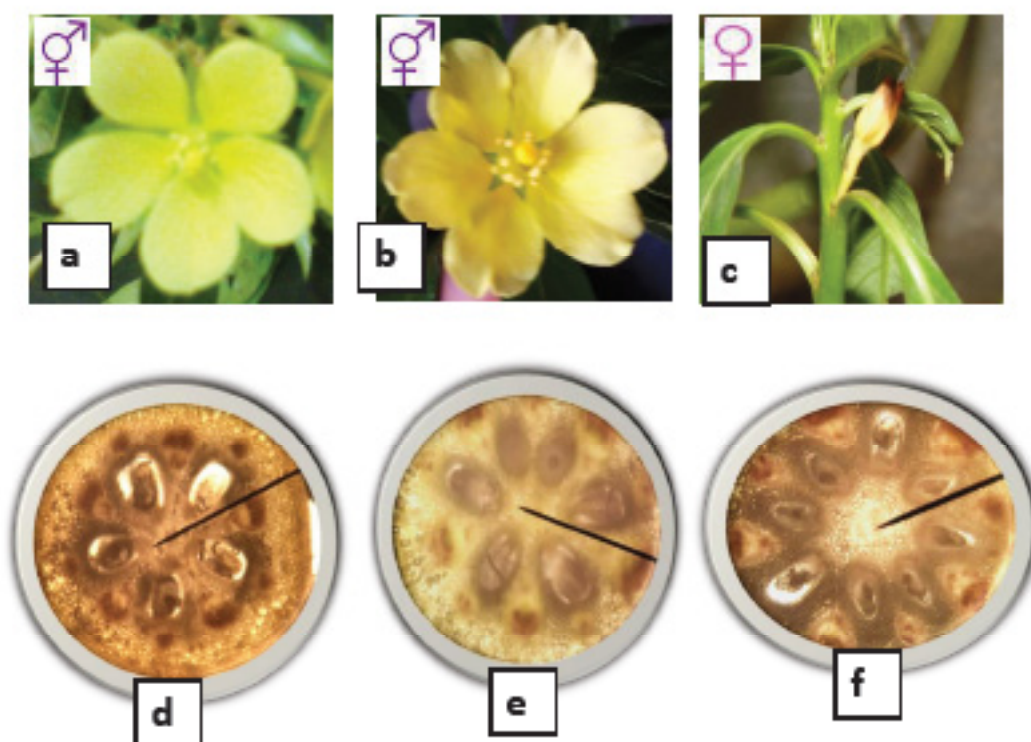


Fig. 6. Flower diversity of *L. stolonifera*, (a) Typical bisexual 5-merous flower with 10 stamens, (b) Bisexual 6-merous flower with 12 stamens, (c) Female flower, (d) 5-locules ovary, (e) 6-locules ovary and (f) 8-locules ovary.

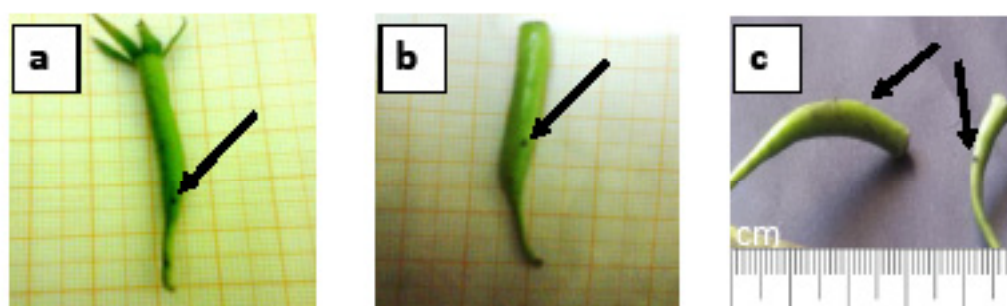


Fig. 7. Fruit diversity in bracteoles position of *L. stolonifera*, (a) Basal in M1, (b) Near base in M4 and (c) Near base or above in M7.

## Discussion

Irrigation and drainage system of River Nile as well as the northern natural lakes in the Nile Delta are highly infested by different communities of aquatic macrophytes, which spread so rapidly and easily fill up the whole of many water bodies,

cause negative effects on the environment, human health and economic development (Fernández et al., 1990; Epstein, 1998; LVEMP, 1999 and Mailu, 2001). The field observation revealed that *Ludwigia stolonifera* is one of the most dominant macrophyte in water courses at least in the area of study.



**Fig. 8. Pneumatophores diversity of *L. stolonifera*, (a&b) Abundant pneumatophores, (c) Occasional pneumatophore and (d) Mixed cluster of conical and cylindrical pneumatophores.**

Clearly, the expansion of *L. stolonifera* in different habitats may threaten plant biodiversity in Egypt. The floristic composition of a community is an indication of species coexistence. Accordingly, the study of the associated plants was necessary. Moreover, floristic composition together with life form represented diagnostic characters to distinguish species especially hydrophytes which have wide geographical and ecological distribution.

In line with our results, Abu Ziada et al. (2008) recognized Poaceae, Chenopodiaceae, Polygonaceae, Amaranthaceae and Cyperaceae to constitute the major part of the floristic composition in their studies on the aquatic vegetation in the north Nile delta. In addition, the two former families were reported to be among the most frequent plant families in Egypt (Shaheen, 2002 and Abd El-Ghani & Fawzy, 2006). Moreover, Poaceae and Chenopodiaceae were found to be the most frequent families containing many weed species in other studies in the tropics (Åfors, 1994; Becker et al., 1998 and Tamado & Milberg, 2000). Furthermore, these families represent the most common in the Mediterranean North African flora (Quézel, 1978). These families are characterized by high number of genera and subsequently they are represented by relatively quite number of species.

The main bulk of the associated species were annuals (29 species) that constituted about 48.3% of

the total flora of the study area. The dominance of annuals could be related to their high reproductive capacity under high level of disturbance (Grime, 1979). The low number of perennials especially in trees (3 species) might be related to the intensive management used in the canals and the continuous mechanical weed control that could affect their vegetative structures.

Based on Raunkiaer's life form system (1937), therophytes were relatively highly represented by 28 species, that constituted about 46.7% of the total number of the associated species, followed by the hydrophytes and helophytes that were represented by 16 species about 26.7%. Similar results were conducted by Abu Ziada et al. (2008) who stated that the therophytes predomintes over other life forms.

*Phragmites australis*, *Eichhornia crassipes* and *Cyperus alopecuroides* were the wide ecological spectrum species in the area of study. These results were also conducted by Khedr & El-Demerdash (1997). Mashaly & El-Amier (2007) recognized these species as indicator species in the irrigation and drainage canals system in egypt. Moreover, Hutchinson (1975) cleared that *Eichhornia crassipes* having high ecological tolerance and large-scale ability to disperse in the fresh and alkaline courses of water.

It is to be noticed that the dense canopy of tall growing species along water edge (e.g. *Phragmites australis*) makes the germination and growth of other species more difficult, this often leads to the reduction in the species diversity. In addition, the human impact, especially the mechanical control of weeds, might be the main reason for the low number of species in association with the brackish morphotype M5 and the fresh to brackish morphotype M4, 6 species and 7 species respectively; this is in line with the result conducted by Shaltout & El-Sheikh (1993).

The high number of species, 32 species (about 53.3% of the collected species), associated with the marine morphotype (M7) may be attributed to the nutrient budget of the saline lakes that is mostly dependent upon the amount of nutrients transported through the waste water drains as the water in these drains contain large amounts of suspended solids (Dowidar & Abdel-Moati, 1983 and Abdel-Moati, 1990). These conditions favor the growth of floating and emergent hydrophytes.

In the present study, a suite of chorotypes such as Cosmopolitan, Paleotropical, Pantropical, Mediterranean, Sudano-zambezian and Saharo-Arabian had been represented. This could be attributed to human impact and the ability of certain phytochoria to penetrate the study area from several adjacent phytogeographical regions (Shalaby, 1995).

Water invaders (Simpson, 1932) are land weeds that grow near water, usually with their root systems below water (helophytes *sensu* Raunkiaer, 1937). These weeds invade the water, either from the bank or from more or less shallow water, some are rooted at the normal water level and their stems and branches spread out over the water. The field observation cleared that form in *Echinochloa crusgalli*. Most of the recorded water species in the present study had relatively maximum occurrence in the wetlands along lake Manzala and Edku and other water courses, belong to Poaceae, Cyperaceae and Polygonaceae (Zahran & Wills, 2003). This might support the abundance of some species like *Persicaria salicifolia*. Njambuya & Triest (2010) showed that *Ludwigia stolonifera*, a native species, outperformed the invasive species in all measures of performance studied under nutrient non-limited conditions. These findings are consistent with the conclusion drawn by Daehler (2003) that invasive

species do not always have higher growth rate and neither do they often competitively outperform co-occurring native species. Some native species have the same attributes as invasive species (Thompson et al., 1995). Our study confirmed that wherever populations of *L. stolonifera* grown, it outperformed other invaders.

In addition, Njambuya & Triest (2010) indicated that *Ludwigia stolonifera* as a creeping emergent macrophyte, had a higher average relative growth rate that could accumulate more biomass. Rejmánková (1992) pointed out that the so-called creeping emergent species accumulate more biomass to above than belowground part. This meets our observations in the field.

Comparison of macro and micro morphological characters among the studied populations of *L. stolonifera* revealed seven morphotypes. The similarities between each pair of the 7 morphotypes was elaborated using Jaccard's Measure. Frankel et al. (1995) and El-sadek & Ayyad (2000) demonstrated that genetic diversity within species gives the opportunity to evolve under changing environments and selection pressure. This may explain the least similarities between the terrestrial wetland morphotype M3 and the other aquatic morphotypes and the relatively high values of similarity between the two fresh to brackish morphotypes M4 & M6 (66.7%) and between the two fresh morphotypes M1 & M2 (53.1%). In addition, this may clarify the response to variations of environmental factors to survival and growth in the heterogeneous environments (Dorken & Barrett, 2004). Similar observation was noticed in the investigation of Khedr & Hegazy (1998) on *Nymphaea lotus* around Manzala lake where this hydrophytic species invaded the rice fields causing a serious problem. However, the high similarity values between marine morphotype M7 and the three morphotypes M1 (fresh morphotype), M4 and M6 (fresh to brackish morphotypes) cleared the high polymorphism of this morphotype, this come in line with Bidak & Marzouk (2005) who observed high degree of acclimation and plasticity accompanied with *L. stolonifera* populations of certain northern lakes especially lake Manzala. This postulate was confirmed by our result on the similarity between each pair of the 7 morphotypes depending on their associated species using the proximity matrix.

The application of CCA in the present data set agreed with Khedr & El-Demerdash (1997) as they showed that water salinity, EC, potassium ions and total phosphates contents were the environmental factors associated with macrophytic distribution, concluded that the distribution of emergent and floating species was best correlated with water salinity, potassium and total phosphates.

The correlation between fresh water morphotypes M1 & M2 and organic matter & turbidity, agreed with the gradual pollution of most of our water resources by the addition of foreign materials from the surroundings. These included organic matter of plant and animal origin, land surface washing, industrial, agricultural and sewage effluents which containing large amount of dissolved ions and high amount of organic and inorganic constituents. This may agree with Elewa & Mahadi (1988).

The results of the CCA biplot is in agreement with the findings conducted by Corre (1985) and Winter (1990) who stated that halophytes have the ability to grow and dominate in habitats with high concentrations of cations, salinity, pH and moisture contents

High cumulative variance (93.5%) of the species-environment correlations suggested an association between the flora and the measured water variables presented in the study area.

*Ludwigia stolonifera* is a fresh water amphibious plant can grow in both terrestrial and aquatic conditions. This feature may be attributed to its phenotypic plasticity, recorded earlier in some species of this genus as in *L. grandiflora* (Ruaux et al., 2009 and Haury et al., 2014a) and *L. octovalvis* (Chauhan & Abugho, 2012).

Hutchinson (1975) demonstrated that the distribution of species of aquatic plants is associated with their large-scale ability to disperse sexual or vegetative propagules that can establish in novel habitats, clonal growth and broad ecological tolerance (Darwin, 1859; Sculthorpe, 1967; Grace, 1993; Les et al., 2003 and Charalambidou & Santamària, 2005); these qualities allow them to occur over a wide range of conditions (Lacoull & Freedman, 2006).

Bar & Ori (2014) indicated that plants are able to change quickly in response to environmental

cues; leaf shape exemplifies this principle well and represents a classic example of developmental plasticity (Arber, 1920; Schmalhausen, 1949; Sculthorpe, 1967; Cook & Johnson, 1968 and Cook, 1968). The variation in the length and shape of leaves among the studied morphotypes ranged from short, spatulate to narrowly elliptic in M3 to relatively long, lanceolate leaves in M2, M4, M5 and M7 morphotypes which reflects the plastic changes in leaf shape that are considered to be an adaptation of the presence or absence of water in the plant's environment (Morisset & Boutin, 1984). This feature come in line with Peng (1988), Lavania et al. (1990) and Scremin-Dias (2009).

Aquatic plants display a remarkable range of reproductive strategies, including diverse sexual systems and means of clonal propagation (Barrett et al., 1993). Reproductive strategies affect the response of populations to environmental heterogeneity (Ronce & Olivieri, 1997; Barrett & Pannell, 1999; Heilbuth et al., 2001 and Crowley & McLetchie, 2002). Moreover, the variation in reproductive traits will influence the ability of populations to colonize and persist in different types of aquatic habitats and these can explain the variation in flower sex of aquatic morphotypes of *L. stolonifera* when compared with the terrestrial morphotype M3.

Bradshaw (1965) and Schlichting (1986) reported that differences in production of seeds is known to occur between populations of many species. This is in line with our study that reflects variations in the fruit morphology and seed productivity/fruit of *L. stolonifera* morphotypes. These may attributed to the variation in the flower which in turn reflect on fruit morphology and seed productivity among the studied morphotypes and come in line with Bidak & Marzouk (2005) who reported variation in seed productivity among *L. stolonifera* populations.

Within the seven morphotypes, the bracteoles are located either at, near the base or higher up on the sides of the capsules which reflect their phenotypic plasticity of the studied morphotypes. This variability may be detected in the same plant. Peng (1989) recorded that variability within the individual of the same plant as in *L. linifolia*, the bracteoles range from (1.5-) 2.5-9 (-13)mm long. Few species including *L. sphaerocarpa* and *L. microcarpa* consistently have shorter bracteoles



(less than 1.5mm long) than other species,

As a member of water living, the morphological constructions of *L. stolonifera* morphotypes vegetative organs especially pneumatophores which contains the ability to strengthen the plant oxygen supply and support the plant up grown may have a kind of plasticity to be able to grow and survive in different habitats as fresh water morphotypes (M1 and M2) to brackish (M4 and M6) to marine morphotypes (M7), while the terrestrial morphotype (M3) has fibrous roots attache to the substrate as it is found along the edges of water bodies. This result is aligned with the observation of Sen (1959) in the floating and land forms of *L. adscendens*, confirmed by Fuchen et al. (2005) who demonstrated that production of pneumatophores in *Ludwigia stolonifera* may differ in response to water depth as populations living in deep water are in need of oxygen as than that living in wetland or shallow water. As well, Haury et al. (2014b) stated that *L. peploides* had a lower percentage of roots compared to *L. grandiflora* present in aquatic habitats.

Clearly, the pollen grains of the studied morphotypes showed notable plasticity on many palynological characters such as the P/E ratio, position of aperture, presence of operculum, presence of crystal-like elements on exine surface and the presence of viscin threads. Habitat diversity was clearly reflected on the recorded plasticity where the morphotype inhabiting the saline habitat can easily differentiated by its planaperturate pollen from others inhabiting fresh water habitats by their angulaperturate pollen. On the same way, the morphotypes inhabiting brackish habitat characterized by long colpus and presence of operculum, similar results were observed by Torabi et al. (2013) who detected various abnormalities and unusual adhesion of pollen grains in saline condition when compared with those in normal conditions.

Jain (1978) cleared that such interspecific variabilities could be of adaptive significance and could affect population fitness, particularly in inhabiting fluctuating and unpredictable environments. In addition, Frankel et al. (1995) and El-sadek & Ayyad (2000) demonstrated that genetic diversity within species gives the oppourtunity to evolve under changing environments and selection pressure. So we can conclude that further cytological and

molecular studies need to elucidate the impact of such genetic diversity on plasticity among *L. stolonifera* morphotypes.

### **Conclusion**

The studied populations of *Ludwigia stolonifera* exhibit high phenotypic plasticity in relation to the different habitats (terrestrial, fresh water, brackish and saline) in the leaf shape and size, the flower characters, number of carpels and stamens, the size and position of bracteoles, the seed productivity per fruit, the abundance of vesicles and the position of aperture. The study revealed that this plasticity is correlated with the habitat diversity and the water characteristics. Accordingly, the associated species shows great dissimilarities among the different morphotypes.

### **References**

- Abd El Ghani, M. and Fawzy, F. (2006) Plant diversity around springs and wells in five oases of the Western Desert, Egypt. *International Journal of Agriculture and Biology*, **8**(2), 249–255.
- Abdel-Moati, A.R. (1990) An input/output flux for lead in a coastal bay off Alexandria region. *Water, Air and Soil Pollution*, **59**, 261–269.
- Abu Ziada, M.E., Mashaly, I.A. and Torky, M. (2008) Ecological studies on the aquatic vegetation in north east Nile Delta. *International Journal of Botany*, **4**(2), 151–163.
- Åfors, M. (1994) Weeds and weed management in small-scale cropping systems in northern Zambia. Valsåtra-Ultuna, Sweden: Crop Production Science, Swedish University of Agricultural Science.
- Amer, W.M., Hamdy, R.S. and Hamed, A.B. (2016) Macro-and micro-morphological variations of *Ludwigia stolonifera* (Guill. & Perr.) P.H. Raven, its taxonomic assessment. 6<sup>th</sup> *International Conference of the Egyptian Botanical Society*, New Horizons in Botanical, Microbiological Research for Sustainable Environment, Cairo, Egypt. 11–12 May.
- APHA (1995) "Standard Methods for the Examination of Water and Wastewater". 19<sup>th</sup> ed. American Public Health Association, Washington D.C.
- Arber, A. (1920) "Water Plants: A Study of Aquatic

- Angiosperms*". Cambridge University Press. Cambridge, UK.
- ASTM D6919-09 (2009) "*Standard Test Method for Determination of Dissolved Alkali and Alkaline Earth Cations and Ammonium in Water and Wastewater by Ion Chromatography*", ASTM International, West Conshohocken, PA.
- ASTM D4327-11 (2011) "*Standard Test Method for Anions in Water by Suppressed Ion Chromatography*", ASTM International, West Conshohocken, PA.
- Bar, M. and Ori, N. (2014) Leaf development and morphogenesis. *Development*, **141**, 4219–4230.
- Barrett, S.C.H., Eckert, C.G. and Husband, B.C. (1993) Evolutionary processes in aquatic plants. *Aquatic Botany*, **44**, 105–145.
- Barrett, S.C.H. and Pannell, J.R. (1999) Metapopulation dynamics and mating-system evolution in plants. In: "*Molecular Systematics and Plant Evolution*", P. Hollingsworth, R. Bateman, R. Gornall (Ed.). Taylor & Francis, London, UK.
- Becker, B., Terrones, F. and Horchler, P. (1998) Weed communities in Andean cropping systems of northern Peru. *Angewandte Botanik*, **72**, 113–130.
- Berk, K.N. (1994). "*Data Analysis with Student SYSTAT Windows Edition Course Technology*", Inc., Cambridge, MA, 595 pp.
- Bidak, L.L.M. and Marzouk, R.I. (2005) Variability among *Ludwigia stolonifera* (Onagraceae) populations in the Nile Delta lakes. *Assiut University Journal of Botany*, **34**(2), 267–287.
- Boulos, L. (1995) "*Flora of Egypt Checklist*". Al Hadara Publishing, Cairo, Egypt.
- Boulos, L. (1999) "*Flora of Egypt*", Vol.1: Azollaceae-Oxalidaceae. Al Hadara Publishing, Cairo, Egypt.
- Boulos, L. (2000) "*Flora of Egypt*", Vol. 2: Geraniaceae-Boraginaceae. Al Hadara Publishing, Cairo, Egypt.
- Boulos, L. (2002) "*Flora of Egypt*", Vol. 3: Verbenaceae-Compositae. Al Hadara Publishing, Cairo, Egypt.
- Boulos, L. (2005) "*Flora of Egypt*", Vol. 4: Alismataceae-Orchidaceae. Al Hadara Publishing, Cairo, Egypt.
- Boulos, L. (2009) "*Flora of Egypt Checklist Revised Annotated Edition*". Al Hadara Publishing, Cairo, Egypt.
- Bradshaw, A.D. (1965) Evolutionary significance of phenotypic plasticity in plants. *Advances in Genetics*, **13**, 115–155.
- Charalambidou, I. and Santamaria, L. (2005) Field evidence for the potential of water birds as dispersers of aquatic organisms. *Wetlands*, **25**, 252–258.
- Chauhan, B.S. and Abugho, S.B. (2012) Phenotypic plasticity of spiny amaranth (*Amaranthus spinosus*) and long fruited primrose-willow (*Ludwigia octovalvis*) in response to rice interference. *Weed Science*, **60**(3), 411–415.
- Cook, C.D.K. (1968) Phenotypic plasticity with particular reference to three amphibious plant species. In: "*Modern Methods in Plant Taxonomy*", V.H. Heywood (Ed.), pp. 97–111. Academic Press London, UK.
- Cook, S.A. and Johnson, M.P. (1968) Adaptation to heterogeneous environments. I. Variation in heterophylly in *Ranunculus flammula* L. *Evolution*, **22**, 496–516.
- Corre, J.J. (1985) Environmental structure and variation in coastal vegetation of the Gulf du Lion (France). *Vegetatio*, **61**, 15–22.
- Crowley, P.H. and McLetchie, D.N. (2002) Trade-offs and spatial life history strategies in classical metapopulations. *American Naturalist*, **159**, 190–208.
- Daehler, C.C. (2003) Performance comparison of co-occurring native and alien invasive plants: implications for conservation and restoration. *Annual Review of Ecology Evolution and Systematics*, **34**, 183–221.
- Darwin, C. (1859) "*On the Origin of Species by Means of Natural Selection, or the Preservation of Favoured Races in the Struggle for Life*". London, UK: John Murray.
- Dorken, M.E. and Barrett, S.C.H. (2004) Phenotypic plasticity of vegetative and reproductive traits in monoecious and dioecious populations of

- Sagittaria latifolia* (Alismataceae), a clonal aquatic plant. *Journal of Ecology*, **92**, 32–44.
- Dowidar, M.N. and Abdel-Moati, R.A. (1983) Distribution of nutrient salts in Lake Manzalah (Egypt). *Rapp P-V Comm. Int. Mer Médit.* **28**, 185–188.
- El-Sadek, L.M. and Ayyad, M. (2000) Genetic diversity as a basic component of biodiversity: Case studies in Egypt. In: "*Plant Systematics for the 21<sup>st</sup> Century*", B. Nordenstam, G. El-Ghazaly, N. Kassas, T. Laurent (Ed.), pp. 239–250. Portland Press, London, UK.
- Elewa, A.A. and Mahadi, H. (1988) Some limnological studies on the Nile water at Cairo, Egypt. *Bulletin of the National Institute of Oceanography and fisheries*, **14**(2), 141–152.
- EPPO, European and Mediterranean Plant Protection Organization, (2004) *Ludwigia peploides* and *L. uruguayensis* (= *L. grandiflora*). EPPO Alert List. European and Mediterranean Plant Protection. (Online). Website <http://www.eppo.org/QUARANTINE/plants/Ludwigia/LUDSS.htm>. (Accessed 12 June 2009).
- Epstein, P. (1998) Weeds bring disease to the east African waterways. *Lancet*, **351**, 577.
- Fernández, O.A., Sutton, D.L., Lallana, V.H., Sabbatini, M.R. and Irigoyan, J.H. (1990) Aquatic weeds problems and management in South and Central America. In: "*Aquatic Weeds: The Ecology and Management of Nuisance Aquatic Vegetation*", A.H. Pieterse, K.J. Murphy (Ed.), pp. 406–425. Oxford University Press, Oxford, UK.
- Frankel, O.H., Brown, A.H.D. and Burdon, J.J. (1995) "*The Conservation of Plant Biodiversity*". Cambridge University Press, Cambridge, UK.
- Fuchen, C., Guoxin, S., Ruowen, C., Hongliang, X. and Min, Z. (2005) Studies on morphological structure of *Jussiaea repens* Linn. and its adaptation to the aquatic habitats. *Journal of Natural Science Nanjing Normal University*, **7**(2), 1–5.
- Gauch, H.G. and Whittaker, R.H. (1981) Hierarchical classification of community data. *Journal of Ecology*, **69**, 537–557.
- Global Invasive Species Database (2009) *Ludwigia peruviana*. website <http://www.invasivespecies.net/database/welcome/disclaimer.asp> (accessed 12 June 2009).
- Grace, J.B. (1993) The adaptive significance of clonal reproduction in angiosperms: An aquatic perspective. *Aquatic Botany*, **44**, 159–180.
- Grime, J.P. (1979) "*Plant Strategies and Vegetation Processes*". John Wiley and Sons. New York, USA.
- Hauray, J., Druel, A., Cabral, T., Paulet, Y., Bozec, M. and Coudreuse, J. (2014a) Which adaptations of some invasive *Ludwigia* spp. (Rosidae, Onagraceae) populations occur in contrasting hydrological conditions in Western France? *Hydrobiologia*, **737**(1), 45–56.
- Hauray, J., Coudreuse, J., Bozec, M. and Barloy, D. (2014b) Biology, ecology and fertility of Water Primroses (*Ludwigia grandiflora* subsp. *hexapetala* and *Ludwigia peploides* subsp. *montevicensis*): Variability and consequences for management in Western France, *4<sup>th</sup> International Symposium on Environmental Weeds and Invasive Plants*. UMR INRA AO ESE, France.
- Heilbuth, J.C., Ilves, K.L. and Otto, S.P. (2001) The consequences of dioecy for seed dispersal: Modeling the seed-shadow handicap. *Evolution*, **55**, 880–888.
- Hill, M.O. (1979) TWINSPAN-A FORTRAN Program for Arranging Multivariate Data in an Ordered Two-Way Table of Classification of Individuals and Attributes. Ithaca: Cornell University.
- Hutchinson, G.E. (1975) "*A Treatise on Limnology: III. Limnological Botany*", p. 660. John Wiley and Sons, New York, London, Sydney and Toronto.
- Jain, S.K. (1978) Inheritance of Phenotypic plasticity in soft chess, *Bromus mollis* L. (*Gramineae*). *Experientia*, **34**, 835–836.
- Khedr, A.A. and El-Demerdash, M.A. (1997) Distribution of aquatic plants in relation to environmental factors in the Nile Delta. *Aquatic Botany*, **56**, 75–86.
- Khedr, A.A. and Hegazy, A.K. (1998) Ecology of the rampant weed *Nymphaea lotus* (L.) Willdenow in natural and rice field habitats of the Nile Delta, Egypt. *Hydrobiologia*, **386**, 119–129.

- Lacoull, P. and Freedman, B. (2006) Environmental influences on aquatic plants in freshwater ecosystems. *Environmental Reviews*, **14**, 89–136.
- Lavania, G.S., Paliwal, S.C. and Gopal, B. (1990) Aquatic vegetation of the Indian subcontinent. In: "*Ecology and Management of Aquatic Vegetation in the Indian Subcontinent*", B. Gopal (Ed.), 1<sup>st</sup> ed, pp. 29–78. Kluwener Academic Publishers, Dordrecht, Boston, London.
- Les, D.H., Crawford, D.J., Kimball, R.T., Moody, M.L. and Landolt, E. (2003) Biogeography of discontinuously distributed hydrophytes: A molecular appraisal of intercontinental disjunctions. *International Journal of Plant Science*, **164**, 917–932.
- LVEMP (1999) Lake Victoria Environmental Management Project. Report of the Second East African Cooperation Ministerial Meeting on Water Hyacinth, August 19, 1999. Report Ref. No. EAC/SR/12/99, AICC, Arusha, Tanzania.
- Mailu, A.M. (2001) Preliminary assessment of the social, economic and environmental impacts of water hyacinth in the Lake Victoria basin and the status of control. In Biological and Integrated Control of Water Hyacinth, *Eichhornia crassipes*. *Proceedings of the Global Working Group for the Biological Control and Integrated Control of Water Hyacinth*, Beijing, China. ACIAR proceedings no. 102, pp. 62–70.
- Mashaly, I. and El-Ameir, Y.A. (2007) Hydrophytic vegetation in the irrigation and drainage canal system of the River Nile in Egypt. *World Applied Sciences Journal*, **2**(1), 49–61.
- Morriset, P. and Boutin, C. (1984) The biosystematic importance of phenotypic plasticity. In: "*Plant Biosystematics*", Grant, W.F. (Ed), pp. 293–306. Academic Press, New York, USA.
- Müller-Dombois, L.D. and Ellenberg, H. (1974) "*Aims and Methods of Vegetation Ecology*". John Wiley and Sons, New York, USA.
- Njambuya, J. and Triest, L. (2010) Comparative performance of invasive alien *Eichhornia crassipes* and native *Ludwigia stolonifera* under non-limiting nutrient conditions in Lake Naivasha, Kenya. *Hydrobiologia*, **656**(1), 221–231.
- Økland, R.H. (1990) Vegetation ecology: Theory, methods and applications with reference to Fennoscandia. *Sommerfettia Supplement*, **1**, 1–216.
- Peng, C.I. (1988) The biosystematics of *Ludwigia* sect. *Microcarpium* (Onagraceae). *Annals of the Missouri Botanical Garden*, **75**, 970–1009.
- Peng, C.I. (1989) The systematics and evolution of *Ludwigia* sect. *Microcarpium* (Onagraceae). *Annals of the Missouri Botanical Garden*, **76**, 221–302.
- Price, T.D., Qvarnström, A. and Irwin, D.E. (2003) The role of phenotypic plasticity in driving genetic evolution. *Proceedings of the Royal Society of Biological Sciences*, **270**(1523), 1433–1440.
- Punt, W., Hoen, P.P., Blackmore, S., Nilsson, S. and Le Thomas, A. (2007) Glossary of pollen and spore terminology. *Review of Palaeobotany and Palynology*, **143**, 1–81.
- Quézel, P. (1978) Analysis of the flora of Mediterranean and Saharan Africa. *Annals of the Missouri Botanical Garden*, **65**, 479–534.
- Raunkiaer, C. (1937) "*The Plant Life Forms and Statistical Plant Geography*". Clarendon Press, Oxford, UK.
- Rejmánková, E. (1992) Ecology of creeping macrophytes with special reference to *Ludwigia peploides* (H. B. K.) Raven. *Aquatic Botany*, **43**, 283–299.
- Ronce, O. and Olivieri, I. (1997) Evolution of reproductive effort in metapopulation with local extinctions and ecological succession. *American Naturalist*, **150**, 220–249.
- Ruau, B., Greulich, S., Haury, J. and Berton, J.P. (2009) Sexual reproduction of two alien invasive *Ludwigia* (Onagraceae) on the middle Loire River, France. *Aquatic Botany*, **90**, 143–148.
- Schlichting, C.D. (1986) The evolution of phenotypic plasticity in plants. *Annual Review of Ecology and Systematics*, **17**, 667–693.
- Schmalhausen, I.I. (1949) "*Factors of Evolution: The Theory of Stabilizing Selection*". University of Chicago Press, Chicago, Illinois.
- Scremin-Dias, E. (2009) Tropical aquatic plants:

- Morphoanatomical adaptations. In: "*Encyclopedia of Tropical Biology and Conservation Management*", K. Del-Claro, V. Rico-Gray (Ed.), Vol. 1, pp. 84–132. Natural History of Tropical Plants. Paris, France: UNESCO/EOLSS.
- Sculthorpe, C.D. (1967) "*The Biology of Aquatic Vascular Plants*". Edward Arnold, London. 610p.
- Sen, D.N. (1959) Ecological studies on aquatic and swampy vegetation of Gorakhpur. *Agra University Journal of Research Science*, 8, 17–29.
- Shaheen, A.M. (2002) Weed diversity of newly formed land on the southern border of Egypt (Eastern and Eastern shores of Lake Nassar). *Pakistan Journal of Biological Sciences*, 5, 602–608.
- Shalaby, M.A. (1995) Studies on plant life at Kafr El Sheikh Province, Egypt. *M.Sc. Thesis*, Kafr El Sheikh, Tanta University, Egypt.
- Shaltout, K.H. and El-Sheikh, M.A. (1993) Vegetation environment relationships along water courses in the Nile Delta region. *Journal of Vegetation Science*, 4, 567–570.
- Simpson, N.D. (1932) A Report on the Weed Flora of the irrigation channels in Egypt. Ministry of Public Work, Government Press, Cairo, 124 pp.
- Täckholm, V. (1974) "*Student's Flora of Egypt*". 2<sup>nd</sup> ed. Cairo University Press, Egypt, Beirut.
- Tamado, T. and Milberg, P. (2000) Weed flora in arable fields of eastern Ethiopia with emphasis on the occurrence of *Parthenium hysterophorus*. *Weed Research*, 40, 507–521.
- Ter-Braak, C.J.E. and Prentice, I.C. (1988) A theory of gradient analysis. *Advances of Ecological Research*, 18, 271–317.
- Ter-Braak, C.J.F. and Smilauer, P. (2002) CANOCO Reference Manual and CANODRAW for Windows User's Guide, Software for Canonical Community Ordination (Version 4.5). Microcomputer Power, Ithaca, New York.
- Thompson, K., Hodgson, J.G. and Rich, T.C.G. (1995) Native and alien invasive plants: more of the same? *Ecography*, 18, 390–402.
- Torabi, F., Majd, A., Enteshari, S., Irian, S. and Nabiuni, M. (2013) Effects of salinity on the development of hydroponically grown borage (*Borago officinalis* L.) male gametophyte. *Notulae Botanicae Horti Agrobotanici*, 41(1), 65–72.
- Ward, D., Olsvig-Whittaker, L. and Lawes, M. (1993) Vegetation-environment relationships in a Negav Desert erosion cirque. *Journal of Vegetation Science*, 4, 83–94.
- Winter, E. (1990) Characteristics and distribution of halophytes at a Jordanean Sultpan. *Flora*, 184, 341–367.
- Zahran, M.A. and Wills, A.J. (1992) "*The Vegetation of Egypt*". Chapman and Hall, London, UK.
- Zahran, M.A. and Willis, A.J. (2003) "*Plant Life in the River Nile in Egypt*". Mars Publishing House Rayadh, Saudi Arabia.
- Zohary, M. (1947) A vegetation map of western Palestine. *Journal of Ecology*, 34, 1–18.
- Zohary, M. (1962) "*Plant life of Palestine, Israel and Jordan*". The Ronald Press Company, New York, USA.

(Received 24/ 9/2018;  
accepted 21/10/2018)

## لودويجيا ستولونيفرا نظرة موضوعية فى المرونة الظاهريه وتنوع الموائل والنباتات المصاحبة

أشرف توفيق سليمان، ريم سمير حمدى و عزة بدر حامد  
قسم النبات والميكروبيولوجي - كلية العلوم - جامعة القاهرة - الجيزة - القاهرة - مصر .

بمتابعة اثنين وثلاثين عشيرة نباتية لنبات المديد (اللودويجيا ستولونيفرا) تم رصد سبعة أنماط مورفولوجية مختلفة تنتمي هذه الأنماط إلى خمسة موائل مختلفة. وكان أكثر النباتات مصاحبة لنبات المديد هي الحجنة وياسنت الماء والسمر الحلو. مثلت النباتات واسعة الانتشار ونباتات الإستوائية للعالم القديم والحديث حوالي 56.7% من مجموع النباتات المصاحبة للمديد. وكذلك مثلت النباتات الحولية ما يقرب من 46.7% ونباتات المائية حوالي 26.7% بينما تواضع تواجد النباتات الأخرى. تم رصد معامل التماثل بين الأنماط السبعة المختلفة معتمدا على 42 من الخصائص المورفولوجية الدقيقة والكبيرة.

ومما هو جدير بالذكر أن نبات المديد للمياة العذبة تأثر بتواجد المادة العضوية ومعامل عكارة الماء بينما تأثر مديد مناطق الأراضي الرطبة بتواجد الكربونات والأمونيا، وتأثر نبات المديد لمناطق المياه المالحة بمعامل التوصيل الكهربائي والملوحة ومحتوى كلوريد الصوديوم. وقد أظهرت الدراسة مرونة مظهرية متباينة لنبات المديد في الموائل المختلفة، وقد رصدت الدراسة ذلك في اختلاف شكل الأوراق وحجمها، وفي الزهرة من زهرة خماسية بها عشرة أسدية إلى زهرة سداسية بها اثني عشرة من الأسدية، وفي حجم وموضع الوريقة القنابية من قاعدية إلى قرب قاعدية أو تحت طرفية، وفي إنتاجية البذور من 60-80 إلى فقط 35-45 بذرة للثمرة، وفي الجذور من 10 جذور للعنقود الواحد إلى فقط ثلاثة جذور، وأخيرا في وضع فتحات حبوب اللقاح من سطحية إلى ذات الزوايا.