

Genetic Differentiation of *Citrullus colocynthis* (L.) Populations Depending on Allozyme Diversity

Shawkat M. Ahmed^{(1,2)#} and Khalid H. Alamer⁽¹⁾

⁽¹⁾ Biology Department, Faculty of Science, Taif University, PO Box 5700, Taif, KSA;

⁽²⁾ Biology Department, Faculty of Education, Ain Shams University, Cairo, Egypt.

CITRULLUS *colocynthis* (L.) is a desert plant having a large history in pharmaceutical industries as an important medicinal plant. Allozyme variation and population genetic structure of *C. colocynthis* in 6 natural populations from western region, Saudi Arabia, were investigated by 6 enzyme systems. The genetic differentiation within and among *C. colocynthis* populations has been revealed by allozyme analysis. Ten loci were monomorphic in all populations, whereas seven loci having two alleles (α -EST-1, β -EST-1, MDH-1, ADH-4, ADH-5, ALO-4 and PRX-2) were polymorphic. The mean allozyme variability indices within *C. colocynthis* populations ($P = 31.17\%$; $A = 1.21$) were lower than those for insect-pollinated species ($P = 34\%$; $A = 2.67$; $H_e = 0.205$) except H_e value (0.415) was higher. The observed mean heterozygosity ($H_o = 0.757$) was higher than the expected mean heterozygosity ($H_e = 0.415$) indicating the existence of natural selection against homozygosity. The Inbreeding coefficient values (F) of polymorphic loci were negative suggesting a significant excess of heterozygosity in the studied populations. The UPGMA dendrogram of 84 individuals of *C. colocynthis* confirmed the notable variability within populations. Differentiation among-population (F_{ST}) recorded 16.2% of the total variation. Low F_{ST} and high genetic distance (D) values suggested a low differentiation among *C. colocynthis* populations. Little gene flow (N_m) was deduced in all populations except ME based on the allozyme data. We suggest that population ME can be recommended for both genetic conservation and breeding programs.

Keywords: *Citrullus*, Allozyme, Population, Heterozygosity, Gene flow.

Introduction

Citrullus colocynthis L. (Cucurbitaceae), is a drought-resistant wild perennial herb that widely found in Africa and Asia. *C. colocynthis* has many medicinal advantages against various diseases such as diabetes II and breast cancer (Grossman et al., 2007 and Huseini et al., 2009). *C. colocynthis* grows fast in the sandy soil regions of Saudi Arabia under changeable environmental conditions (Chaudhary & AL-jowaid, 1999). Although the wide distribution of *C. colocynthis*, our knowledge about its genetic diversity is very poor. Loss of genetic variation decreases the ability of wild species to survive fluctuations in climate, diseases, pollutants and natural enemies (Frankham, 1995). Genetic diversity and good management of its natural populations are needed for more adaptation and improvement in future. Isozymes are useful in the field of biochemical

genetics and population genetics as biochemical markers (Wendel & Weeden, 1989). Isozymes examined genetic variation and phylogenetic relationships in several *Citrullus* spp. by Navot & Zamir (1987), Biles et al. (1989) and Navot et al. (1990), but most of them were monomorphic. However they will be useful and accurate when we examine more accessions and individuals than some DNA markers (Klaas, 1998; Levi et al., 2001b; Ritschel et al., 2004; Solmaz et al., 2010 and Minsart et al., 2011).

Few studies described the genetic variability and population structure in *C. colocynthis* using isozymes. The aim of this work is to estimate the genetic differentiation within and among six natural populations of *C. colocynthis* from western region of Saudi Arabia, based on isozyme polymorphism. This could be useful for conservation and breeding purposes.

#Corresponding author email: shamahmoh@gmail.com, Postal code: 5700, Phone +966535923238, Fax +966-2-7256620.

Edited by: Prof. Dr. Soliman A. Haroun, Faculty of Science, Kafr El-Sheikh University, Kafr El-Sheikh, Egypt.

DOI: 10.21608/ejbo.2018.3595.1170

©2018 National Information and Documentation Center (NIDOC)

Materials And Methods

Plant materials

Leaves of eighty four individuals of *C. colocythis* (14 individuals in each of six populations studied) were collected for assessment of genetic variability. The six populations were collected from six sites from the western region of Saudi Arabia (Table 1).

Isozymes

The examined isozymes were: Alcohol dehydrogenase (ADH); (E.C. 1.1.1.1), aldehyde oxidase (ALO); (E.C.1.2.3.1), α - and β -esterases (EST); (E.C.3.1.1.1), peroxidase (PRX); (E.C.1.11.1.7) and malate dehydrogenase (MDH); (E.C.1.1.1.37). Extraction of isozyme was carried out by homogenizing 1g of fresh leaves in 1ml extraction buffer (1M Tris-HCl, pH 8.8) then centrifuged at 10000rpm and the supernatant was separated using native-polyacrylamide gel electrophoresis method (Stegemann et al., 1985). Protocols of Scandalios (1964) for α - and β -EST, Wendel & Weeden (1989) for ALO, Weeden & Wendel (1990) for ADH, Jonathan & Wendell (1990) for MDH and Heldt (1997) for PRX were used for isozyme staining.

Data analysis

Parameters for detecting the within-populations genetic variation and the inbreeding coefficient; Wright's F [$F = (1 - H_o/H_e)$], were estimated as described by Hamrick & Godt (1989) and Nei (1973) (Tables 3 and 4). Negative F indices refers to tendency to heterozygosity whereas positive values indicate an inbreeding system of mating. Levels of significance among populations for each

parameter were determined by t-test (Varghese et al., 1999). Parameters of populations genetic structure; F_{IS} , F_{IT} and F_{ST} , were investigated by Wright's F -statistics (Wright, 1965). Gene flow; Nm , was calculated by Wright's (1951) equation with modification by Crow & Alok (1984): $F_{ST} = 1/(4N_m\alpha + 1)$ where $\alpha = [n/(n-1)]^2$ and n is the populations number. Isozyme frequency data were used to calculate genetic distance estimates according to Nei (1973). To discern the genetic variability within populations, each band of 84 individuals was coded as 1 for presence or 0 for absence. Clustering was performed using UPGMA procedure and represented in a phenogram by using SAHN and TREE modules, respectively, and it was also performed for studying the relationships among populations. The computer program NTSYS-pc 2.2 was used in all previous operations (Rohlf, 1998).

Results

Allele diversity

Polymorphisms were observed in all the six systems analyzed. A total of 31 alleles were detected among the 24 loci from 6 enzyme systems evaluated. The frequencies of them are demonstrated in Table 2. Ten loci (α -EST-2, α -EST-3, α -EST-4, α -EST-5, β -EST-4, β -EST-5, MDH-2, ADH-2, ADH-3 and PRX-3) were monomorphic in all populations, whereas seven loci having two alleles (α -EST-1, β -EST-1, MDH-1, ADH-4, ADH-5, ALO-4 and PRX-2) were polymorphic in at least one population. The two loci; β -EST-3 and PRX-1 were only detected in populations TA and JE, respectively.

TABLE 1. Code, sample size (N) and sites of six natural populations of *C. colocythis* from western region of Saudi Arabia.

Population	Pop. code	N	Geographic origin	Latitude North	Longitude East	Altitude (m)	Rainfall (mm/year)
Taif	TA	14	Taif city	21°16'N	40°24'E	1672	119
Jeddah	JE	14	170 km W of Taif	21°32'N	39°11'E	21	53.5
Mecca	ME	14	63 km W of Taif	21°25'N	39°49'E	300	111.8
Rabigh	RA	14	222 km N W of Taif	22°47'N	39°2'E	13	44
Al Khurmah	KH	14	120 km E of Taif	21°54'N	42°1'E	1020	390
Al Bahah	BA	14	250 km S of Taif	20° 24' N	41° 57'E	2300	138.1

TABLE 2. Estimated allele frequencies at 24 loci in six population of *C. colocynthis*.

Locus	Alleles	Population					
		JE	TA	ME	RA	KH	BA
α -EST-1	a	0.50	0.00	0.50	0.00	0.50	0.50
	b	0.50	0.00	0.50	0.00	0.50	0.50
α -EST-2	a	1.00	1.00	1.00	1.00	1.00	1.00
α -EST-3	a	1.00	1.00	1.00	1.00	1.00	1.00
α -EST-4	a	1.00	1.00	1.00	1.00	1.00	1.00
α -EST-5	a	1.00	1.00	1.00	1.00	1.00	1.00
β -EST-1	a	0.50	0.50	1.00	0.50	0.50	1.00
	b	0.50	0.50	0.00	0.50	0.50	0.00
β -EST-2	a	1.00	1.00	0.00	0.00	0.00	0.00
β -EST-3	a	0.00	1.00	0.00	0.00	0.00	0.00
β -EST-4	a	1.00	1.00	1.00	1.00	1.00	1.00
β -EST-5	a	1.00	1.00	1.00	1.00	1.00	1.00
MDH-1	a	0.50	0.50	0.50	0.63	0.67	0.67
	b	0.50	0.50	0.50	0.37	0.33	0.33
MDH-2	a	1.00	1.00	1.00	1.00	1.00	1.00
ADH-1	a	0.00	0.00	1.00	1.00	0.00	1.00
ADH-2	a	1.00	1.00	1.00	1.00	1.00	1.00
ADH-3	a	1.00	1.00	1.00	1.00	1.00	1.00
ADH-4	a	0.50	0.50	0.50	0.50	0.50	0.50
	b	0.50	0.50	0.50	0.50	0.50	0.50
ADH-5	a	0.36	0.00	0.63	0.86	0.93	1.00
	b	0.64	1.00	0.37	0.14	0.07	0.00
ALO-1	a	1.00	1.00	1.00	1.00	1.00	0.00
ALO-2	a	1.00	1.00	1.00	1.00	1.00	0.00
ALO-3	a	1.00	1.00	1.00	1.00	1.00	0.00
ALO-4	a	0.71	0.93	1.00	0.93	1.00	0.89
	b	0.29	0.07	0.00	0.07	0.00	0.11
PRX-1	a	1.00	0.00	0.00	0.00	0.00	0.00
PRX-2	a	0.79	0.00	1.00	0.00	0.96	0.00
	b	0.21	0.00	0.00	0.00	0.04	0.00
PRX-3	a	1.00	1.00	1.00	1.00	1.00	1.00

Genetic variation within populations

Parameters of genetic variability showed considerable level of genetic variation within populations (Table 3). A ranged from 1.14 in population ME to 1.30 in population KH with a mean of 1.21 alleles per locus, whereas the mean of A_p was 1.72, ranging from 1.56 in populations KH to 1.97 in population ME. P ranged from 25.0% in population TA to 35.3% in population BA with a mean of 31.17%. H_o ranged from 0.636 in population RA to 0.938 in population ME, whereas the range of H_e was from 0.358 in population KH to 0.492 in population ME. The observed heterozygosities were higher than those expected in all populations. Obviously population ME possessed the highest level of genetic variation H_e , while populations RA and KH had rather lower levels of genetic variation. The mean observed heterozygosity was higher than that expected, with values of 0.757 and 0.415, respectively. The UPGMA dendrogram (Fig. 1) obtained using the 33 isozyme bands scored in the 84 individuals, showed a notable variability within populations examined. At coefficient 0.59, ten groups were formed, five of them; G1, G3, G4, G5 and G6, corresponded to populations JE, RA, BA,

ME and KH, respectively. Most of the individuals of the same population grouped together, except population TA, which presented lower similarity values among individuals, representing the higher genetic diversity within population.

The F values of all loci were lower than zero. One exception; locus PRX-2 in population KH was greater than zero reflecting a deficiency of heterozygosity (Table 4). The Inbreeding coefficient ranged from -0.902 in population ME to -0.578 in population RA with mean value of -0.602 . Obviously, *C. colocynthis* populations revealed an excess of heterozygosity.

Genetic variation among populations

F_{IS} and F_{IT} estimates were generally negative. F_{IS} ranged from -0.958 for population ME to -0.60 for population RA, and averaged -0.788 , indicating significant excess of heterozygosity in the populations, which was in accordance with the values of F mentioned above. F_{ST} ranged from 0.008 for population ME to 0.306 for population KH, with a mean of 0.162, indicating that 16.2% of the total genetic variation exists among populations.

TABLE 3. Genetic variability estimates within *C. colocythis* populations.

Population	P	A	A _p	H _o	H _e
JE	31.8	1.27	1.85	0.816	0.458
TA	25.0	1.20	1.69	0.785	0.408
ME	33.3	1.14	1.97	0.938	0.492
RA	26.3	1.16	1.58	0.636	0.367
KH	35.0	1.30	1.56	0.647	0.358
BA	35.3	1.18	1.69	0.720	0.409
Among-population mean	31.17	1.21	1.72	0.757	0.415
SE	1.81	0.03	0.06	0.047	0.021
<i>t</i> -value	17.2***	46.7***	26.6***	16.2***	19.6***

-P (<0.99) percentage of polymorphic loci; *A* average number of alleles per locus; *A_p* average effective number of alleles per locus; *H_o* observed heterozygosity; *H_e* expected heterozygosity under Hardy–Weinberg equilibrium; *F* Fixation Index (Inbreeding Coefficient); SE the standard error.

-ns *P* >0.05; ** *P* <0.01; *** *P* <0.001.

TABLE 4. Inbreeding coefficient values (F) for polymorphic loci in *C. colocythis* populations.

Locus	JE	TA	ME	RA	KH	BA
α-EST-1	-1	-	-1	-	-1	-1
β-EST-1	-1	-1	-	-1	-1	-
MDH-1	-1	-1	-1	-0.609	-0.516	-0.516
ADH-4	-1	-1	-1	-1	-1	-1
ADH-5	-0.540	-	-0.609	-0.203	-0.077	-
ALO-4	-0.383	-0.077	-	-0.077	-	-0.007
PRX-2	-0.295	-	-	-	0.079	-
Mean	-0.745	-0.769	-0.902	-0.578	-0.586	-0.631

Average genetic distance (*D*) values ranged from 0.240 to 0.468 with a mean of 0.325 (Table 5). Consequently, Nei's genetic distances (*D*) and geographic distances were estimated among pairs of *C. colocythis* populations as illustrated in Table 6. On the basis of these genetic distance values, UPGMA cluster analysis showed the genetic relationship of the populations studied through a dendrogram (Fig. 2). Two principal groups were found in the cluster analysis. The first cluster was made up of population JE, TA and BA and the second was formed by the other populations. It appeared that populations of closest geographic proximity could not be clustered together. This

was also confirmed statistically by correlation analysis on Nei's genetic distance and geographic distance ($r = -0.013$; $p = 0.980$). The gene flow (N_m) values ranged between 0.57 and 2.35 in all populations except that of population ME (Table 5). The genetic uniqueness of each population was estimated by F_{ST} , *D* and N_m (Table 5). Lower values of F_{ST} , *D* and higher N_m were obtained for the analyses of population ME (0.008, 0.243, 31.0) than those for all 6 populations (0.162, 0.325, 6.83), respectively. It could be concluded that population ME had some unique genetic features and being suitable for conservation process.

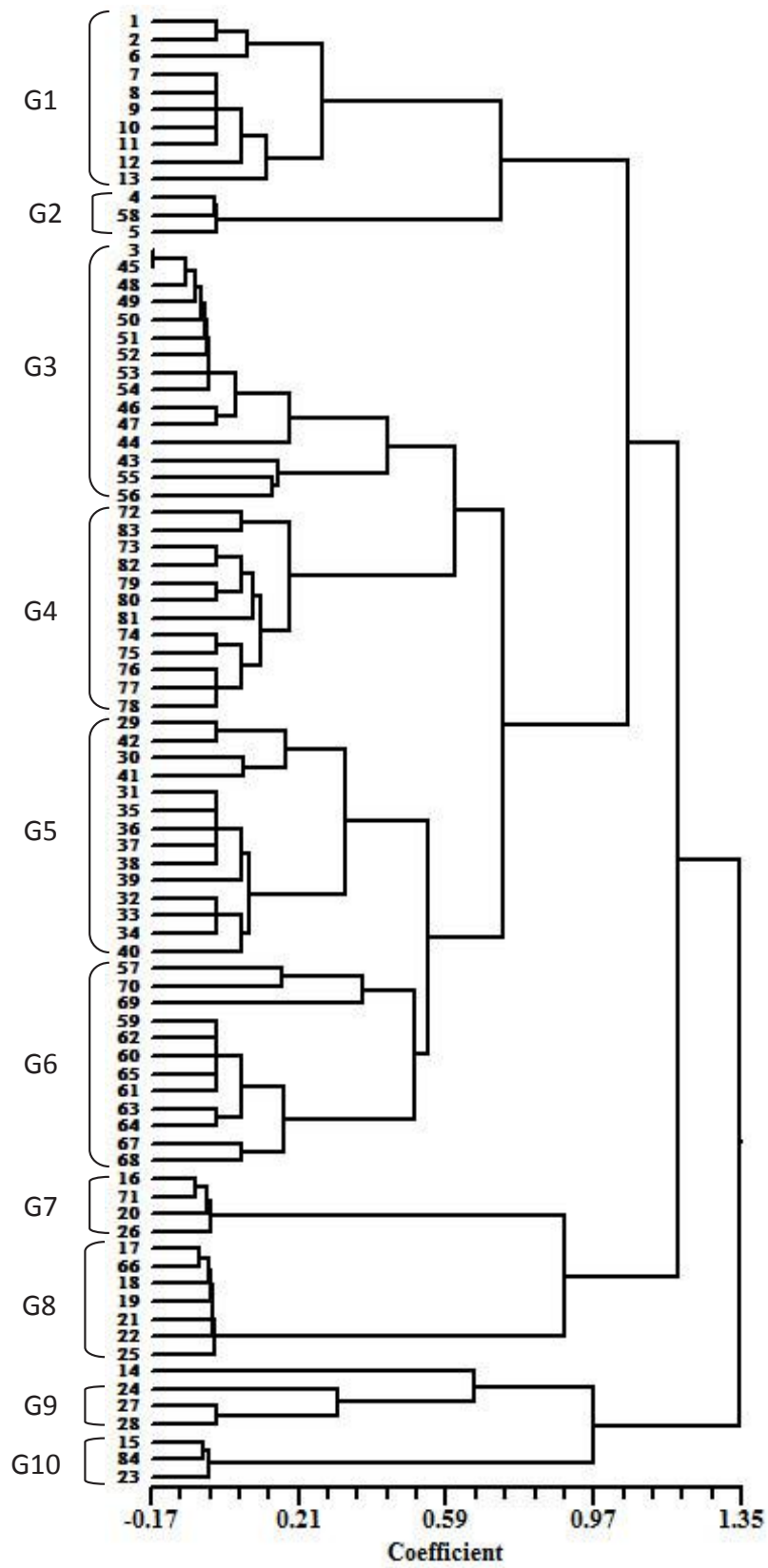


Fig. 1. The dendrogram of 84 individuals of *C. colocynthis* generated by UPGMA analysis.

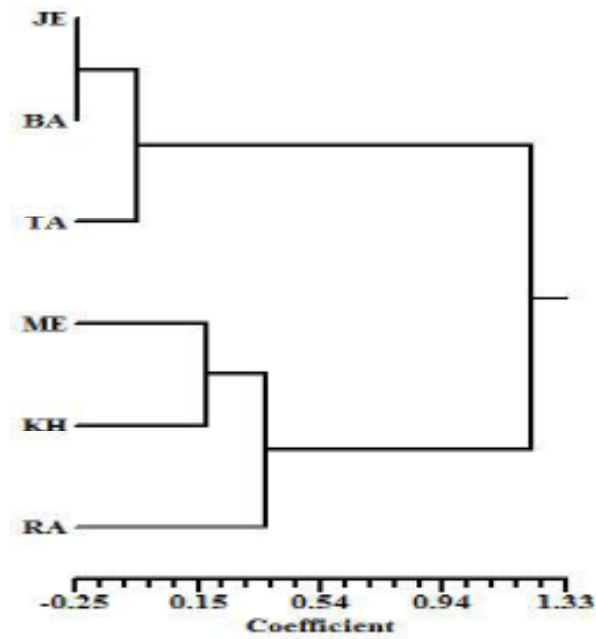


Fig. 2. The UPGMA dendrogram of the six populations of *C. colocythis* based on the Nei's genetic distances.

TABLE 5. F-statistics, average Nei's genetic distance and gene flow values in *C. colocythis* populations.

Pop.	F-statistics						Genetic distance	Gene flow
	H_I	H_S	H_T	F_{IS}	F_{IT}	F_{ST}	D	N_m
JE	0.753	0.443	0.490	-0.699	-0.537	0.096	0.354	2.35
TA	0.754	0.394	0.500	-0.914	-0.508	0.212	0.404	0.93
ME	0.971	0.496	0.500	-0.958	-0.942	0.008	0.243	31.0
RA	0.544	0.340	0.426	-0.600	-0.277	0.202	0.240	0.99
KH	0.540	0.293	0.422	-0.843	-0.279	0.306	0.242	0.57
BA	0.638	0.372	0.436	-0.715	-0.463	0.147	0.468	1.45
Mean	0.700	0.389	0.462	-0.788	-0.501	0.162	0.325	6.21
SE	0.067	0.029	0.016	0.057	0.099	0.042	0.040	4.96
t-test	10.52***	13.16***	29.76***	-13.904***	-5.038**	3.845*	8.098***	1.252 ^{ns}

- F_{IS} inbreeding coefficient within population; F_{IT} total inbreeding; F_{ST} genetic diversity among populations; D average genetic distance; N_m gene flow calculated by Wright's model based on F_{ST} ; SE the standard error.

-ns $P > 0.05$; ** $P < 0.01$; *** $P < 0.001$.

TABLE 6. Geographical distances (km) (above diagonal) and Nei's genetic distances (below diagonal) among *C. colocythis* populations.

	JE	TA	ME	RA	KH	BA
JE		133	65	169	297	272
TA	0.241		68	215	166	187
ME	0.279	0.405		178	232	227
RA	0.354	0.297	0.133		324	400
KH	0.205	0.360	0.108	0.155		220
BA	0.692	0.715	0.288	0.262	0.381	

Discussion

Considerable genetic variation in the six populations of *C. colocynthis* was detected using allozymes, although many researchers predicted low levels of genetic diversity in species having small numbers of individuals and limited ranges (Drury, 1974). The mean allozyme variability indices of *C. colocynthis* within populations ($P=31.17\%$; $A=1.21$) were lower than those of Hamrick & Godt (1997) for insect-pollinated species ($P=34\%$; $A=2.67$; $H_e=0.205$) except H_e value (0.415) was higher. The two P and A estimates were also much lower than those of the four cucurbits; *Cucurbita argyrosperma* ssp. *sororia* (100% - 2.08), *Cucurbita argyrosperma* ssp. *argyrosperma* (93% - 2.5), *Cucurbita moschata* (97% - 2.06) and *Cucurbita pepo* (92% - 2.08) (Montes-hernandez & Eguiarte, 2002). Whereas, our P , A and H_e values were higher than those of *Cucurbita pepo* and *Cucurbita maxima* (Decker-walters et al., 1990), *Cucumeropsis mannii* and *Lagenaria siceraria* (Koffi et al., 2008 and 2009). In all populations the values of H_o were higher than H_e reflecting excess in heterozygosity that was also confirmed by negative inbreeding coefficient (F) values for polymorphic loci. These results conflicted the hypothesis of Biles et al. (1989) and Levi et al. (2001a) that the genus *Citrullus* had a narrow genetic basis, but they were in consistent with Mujaju & Nybom (2011) and Shaik et al. (2015) who found that the diversity was higher in the wild species *C. colocynthis* than *C. lanatus*. This outcome confirmed the hypothesis that the species widely distributed, as *C. colocynthis*, have a higher genetic diversity percentage (H_e) than geographically limited species (Hamrick et al., 1992). The greater allelic diversity within populations suggesting elimination of homozygotes from *C. colocynthis* populations by natural selection and possessing either mating systems characterized by a mixture of self-fertilization and outcrossing (Schoen & Clegg, 1984) or a mode of insect-mediated crosspollination which promoted random mating (Wright, 1951). Previous investigations showed that some species of family Cucurbitaceae had mostly an outcrossing mating system that maintained genetic variation within their populations (Costich & Meagher, 1992 and Montes-hernandez & Eguiarte, 2002).

The F_{IS} and F_{IT} values were negative and revealed an excess of heterozygosity for all

populations. The genetic differentiation among *C. colocynthis* populations ($F_{ST}=0.162$) was similar to that detected for the animal-pollinated plants ($F_{ST}=0.187$) (Hamrick, 1989), but lower than the averages for insect-pollinated species ($F_{ST}=0.245$) and wind-pollinated species ($F_{ST}=0.238$) (Crawford et al., 2001). Similar results were obtained by Koffi et al. (2008 and 2009) who recorded low level of genetic variation among accessions and cultivars of *Cucumeropsis mannii* and *Lagenaria siceraria* compared to the genetic diversity within them. This was in harmony with Hamrick & Godt (1996) and Nybom (2004) who mentioned that species with outcrossing mating system have lower genetic differentiation among populations compared with systems of selfing mating. There was no significant correlation between geographic distance and genetic distance (D). Estimates of gene flow indicated low levels of migration among *C. colocynthis* populations except for population ME. Estimate of $Nm>4$ suggests that gene flow in population ME was enough to face the effects of genetic drift (Kang & Chung, 1997). The extensive gene flow could be attributed to its biological properties, especially its seed dispersal by insects (Dane et al., 2007). However, it appeared that genetic drift might have occurred in other populations which recorded fewer estimates of Nm .

Conclusions

Allozyme markers were powerful to reveal variability in *C. colocynthis*. The mean estimate of F_{ST} (16.2%) indicated that up to 83% of the total genetic variation existed within *C. colocynthis* populations. Therefore, we suggest fewer populations but more individuals within populations for its breeding work. The lack of significant correlation between genetic distance and geographic distance suggested that when sampling populations, it is not necessary to arrange sites identically within the area under study and the population must be selected only if it has high genetic variation. Furthermore, analyses of genetic uniqueness demonstrated that population ME is genetically unique and should also be paid additional interest for conservation purposes.

References

- Biles, C.L., Martyn, R.D. and Wilson, H.D. (1989) Isozymes and general proteins from various

- watermelon cultivars and tissue types. *Hort. Science*, **24**, 810-812.
- Chaudhary, S.A. and Al-Jowaid, A.A. (1999) "Vegetation of the Kingdom of Saudi Arabia". National Agriculture and Water Research Center, Ministry of Agriculture and Water, Riyadh, KSA.
- Costich, D.E. and Meagher, T.R. (1992) Genetic variation in *Ecballium elaterium*: Breeding system and geographic distribution. *Journal of Evolutionary Biology*, **5**, 589-601.
- Crawford, D.J., Ruiz, E., Stuessy, T.F., Tepe, E., Aqveque, P., Gonzalez, F., Jensen, R.J., Anderson, G.J., Bernardello, G., Baeza, C.M., Swenson, U. and Silva, M. (2001) Allozyme diversity in endemic flowering plant species of the Juan Fernandez Archipelago, Chile: Ecological and historical factors with implications for conservation. *American Journal of Botany*, **88**(12), 2195-2203.
- Crow, J.F. and Aloki, K. (1984) Group selection for a polygenic behavioral trait: estimating the degree of population subdivision. *Proceedings of the National Academy of Sciences of the United States of America* **81**, 6073-6077.
- Dane, F., Liu, J. and Zhang, C. (2007) Phylogeography of the bitter apple, *Citrullus colocynthis*. *Genetic Resources and Crop Evolution*, **54**, 327-336.
- Decker-Walters, S.D., Walters, T.W., Posluszny, U. and Kevan, P.G. (1999) Genealogy and gene flow among annual domesticated species of *Cucurbita*. *Canadian Journal of Botany*, **68**, 782-789.
- Drury, W.H. (1974) Rare species. *Biological Conservation*, **6**, 162-169.
- Frankham, R. (1995) Conservation genetics. *Annual Review of Genetics*, **29**, 305-327.
- Grossman, S., Dovrat, S., Gottlieb, H.E. and Bergman, M. (2007) Growth inhibitory activity of cucurbitacin glucosides isolated from *Citrullus colocynthis* on human breast cancer cells. Tannin-Spitz T. *Biochemical Pharmacology*, **73**(1), 56-67.
- Hamrick, J.L. (1989) Isozymes and the analysis of genetic structure in plant populations. In: "Advances in Plant Sciences Series", pp. 87-105. DE Soltis and PS Soltis (Ed.), Dioscorides Press, Portland, Oregon, USA.
- Hamrick, J.L. and Godt, M.J.W. (1989) Allozyme diversity in plant species. In: "Plant Population Genetics, Breeding, and Genetic Resources", A.H.D. Brown, M.T. Clegg, A.L. Kahler and B.S. Weir (Ed.), pp. 43-63. Sinauer Associates Inc., Massachusetts, USA.
- Hamrick, J.L. and Godt, M.W. (1996) Conservation genetics of endemic plant species. In: "Conservation Genetics: Case Studies from Nature", J.C. Avise and J.L. Hamrick (Ed.), pp. 281-304. Chapman and Hall, New York, USA.
- Hamrick, J.L. and Godt, M.J.W. (1997) Effects of life history traits on genetic diversity in plant species. In: "Plant Life Histories-ecology, Phylogeny and Evolution", J. Silvertown, M. Franco and J.L. Harper (Ed.), pp. 102-118. Cambridge University Press, Cambridge, UK.
- Hamrick, J.L., Godt, M.J.W. and Sherman-Broyles, S. (1992) Factors influencing levels of genetic diversity in wood plant species. *New Forest*, **6**, 95-24.
- Heldt, W.H. (1997) A leaf cell consists of several metabolic compartments. "Plant Biochemistry and Molecular Biology". Oxford Univ. Press, Oxford, UK.
- Huseini, H.F., Darvishzadeh, F., Heshmat, R., Jafariazar, Z. and Raza, L.M. (2009) The clinical investigation of *Citrullus colocynthis* (L.) Schrad. fruit in treatment of Type II diabetic patients: A randomized, double blind, placebo-controlled clinical trial. *Phytotherapy Research*, **23**(8), 1186-1189.
- Jonathan, F.W. and Wendell, N.F. (1990) Visualization and interpretation of plant allozyme. In: "Allozymes in Plant Biology", D.E. Sdtis, and P.S. Sottis (Ed.), pp. 5-45. Champan and Hall Press, London, UK.
- Kang, S.S. and Chung, M.G. (1997) Genetic variation and population structure in Korean endemic species: IV. *Hemerocallis hakuunensis* (Liliaceae). *Journal of Plant Sciences*, **110**, 209-217.
- Klaas, M. (1998) Applications and impact of molecular markers on evolutionary and diversity studies in *Allium*. *Plant Breeding*, **117**, 297-308.
- Koffi, K.K., Gbotto, A.A., Malice, M., Dje, Y., Bertin, P., Baudoin, J. and Zoro Bi, I.A. (2008) Morphological and allozyme variation in a collection of *Cucumeropsis mannii* Naudin (Cucurbitaceae)

- from Cote d'Ivoire. *Biochemical Systematics and Ecology*, **36**, 777-789.
- Koffi, K.K., Anzara, G.K., Malice, M., Djè, Y., Bertin, P., Baudoin, J. and Zoro Bi, I.A. (2009) Morphological and allozyme variation in a collection of *Lagenaria siceraria* (Molina) Standl. from Côte d'Ivoire. *Biotechnology, Agronomy, Society and Environment*, **13**(2), 257-270.
- Levi, A., Thomas, C.E., Keinath, A.P. and Wehner, T.C. (2001a) Genetic diversity among watermelon (*Citrullus lanatus* and *Citrullus colocynthis*) accessions. *Genetic Resources and Crop Evolution*, **48**(6), 559-566.
- Levi, A., Thomas, C.E., Wehner, T.C. and Zhang, X. (2001b) Low genetic diversity indicates the need to broaden the genetic base of cultivated watermelon. *Hort. Science*, **36**(6), 1096-1101.
- Minsart, L.A., Dje, Y., Baudoin, J.P., Jacquemart, A.L. and Bertin, P. (2011) Set up of simple sequence repeat markers and first investigation of the genetic diversity of West-African watermelon (*Citrullus lanatus* ssp. *vulgaris* oleaginous type). *Genetic Resources and Crop Evolution*, **58**(6), 805-814.
- Montes-Hernandez, S. and Eguiarte, L.E. (2002) Genetic structure and indirect estimates of gene flow in three taxa of *Cucurbita* (Cucurbitaceae) in western Mexico. *American Journal of Botany*, **89**(7), 1156-1163.
- Mujaju, C. and Nybom, H. (2011) Local-level assessment of watermelon genetic diversity in a village in Masvingo Province, Zimbabwe: Structure and dynamics of landraces on farm. *African Journal of Agricultural Research*, **6**, 5822-5834.
- Navot, N. and Zamir, D. (1987) Isozyme and seed protein phylogeny of the genus *Citrullus* (Cucurbitaceae). *Plant Systematics and Evolution*, **156** (1-2), 61-67.
- Navot, N., Sarfatti, M. and Zamir, D. (1990) Linkage relationships of genes affecting bitterness and flesh color in watermelon. *Journal of Heredity*, **81**, 162-165.
- Nei, M. (1973) Analysis of gene diversity in subdivided populations. *Proceedings of the National Academy of Sciences of the United States of America*, **70**, 3321-3323.
- Nybom, H. (2004) Comparison of different nuclear DNA markers for estimating intraspecific genetic diversity in plants. *Molecular Ecology*, **13**, 1143-1155.
- Ritschel, P.S., Lins, T.C., Tristan, R.L., Buso, G.S., Buso, J.A. and Ferreira, M.E. (2004) Development of microsatellite markers from an enriched genomic library for genetic analysis of melon (*Cucumis melo* L.). *BMC Plant Biology*, **4**(1), 9-22.
- Rohlf, F.J. (1998) *NTSYSpc: Numerical Taxonomy and Multivariate Analysis System*, version 2.02. Exeter Software, New York, USA.
- Scandalios, J.C. (1964) Tissue-specific allozyme variations in maize. *Journal of Heredity*, **55**, 281-285.
- Schoen, D.J. and Clegg, M.T. (1984) Estimation of mating system parameters when outcrossing events are correlated. *Proceedings of the National Academy of Sciences of the United States of America*, **81**, 5258-5262.
- Shaik, R.S., Gopurenko, D., Urwin, N.A.R., Burrows, G.E., Lepschi, B.J. and Weston, L.A. (2015) Population genetics of invasive *Citrullus lanatus*, *Citrullus colocynthis* and *Cucumis myriocarpus* (Cucurbitaceae) in Australia: Inferences based on chloroplast and nuclear gene sequencing. *Biological Invasions*, **17**, 2475-2490.
- Solmaz, I., Sari, N., Aka-Kacar, Y. and Yalcin-Mendi, N.Y. (2010) The genetic characterization of Turkish watermelon (*Citrullus lanatus*) accessions using RAPD markers. *Genetic Resources and Crop Evolution*, **57**(5), 763-771.
- Stegemann, H., Afify, A.M.R. and Hussein, K.R.F. (1986) Identification of date (*Phoenix dactylifera*) cultivars by protein patterns. *Phytochemistry*, **26**(1), 149-153.
- Varghese, M., Edwards, M.A. and Hamrick, J.L. (1999) Genetic variation within two subspecies of *Acacia nilotica*. *Forest Genetics*, **6**(4), 221-228.
- Weeden, N.F. and Wendel, J.F. (1990) Genetics of plant isozymes. In: *Isozymes in Plant Biology*, D.E. Soltis and P.S. Soltis (Ed.), pp. 46-72. Chapman and Hall Press, London, UK.
- Wendel, J.F. and Weeden, N.F. (1989) Visualization and

interpretation of plant allozymes. In: "Allozymes in Plant Biology, Advances in Plant Sciences", Soltis, D.E., Soltis, P.S. (Ed.), series 4, pp. 5-45. Dioscorides Press, Portland, OR, USA.

Wright, S. (1965) The interpretation of population structure by F-statistics with special regard to systems of mating. *Evolution*, **19**, 395-420.

Wright, S. (1951) The genetical structure of populations. *Annals of Eugenics*, **15**, 323-354.

(Received 19/ 4/2018;
accepted 25/ 7/2018)

التمایز الوراثي لعشائر السيتروولوس كولوسينثيس (الحنظل) اعتمادا على تنوع أليلات المشابهات الإنزيمية

شوكت محمود احمد⁽²⁻¹⁾ و خالد حسين العامر⁽¹⁾

⁽¹⁾ قسم الاحياء - كلية العلوم - جامعة الطائف - السعودية و ⁽²⁾ قسم العلوم البيولوجية والبيولوجية - كلية التربية - جامعة عين شمس - القاهرة - مصر.

سيتروولوس كولوسينثيس (الحنظل) هو نبات صحراوي له تاريخ كبير في الصناعات الدوائية باعتباره أحد النباتات الطبية الهامة. وقد تم بحث اختلاف الأليلات المنتجة للمشابهات الإنزيمية وكذلك التركيب الوراثي العشائري داخل نبات الحنظل من خلال 6 عشائر طبيعية تم تجميعها من المنطقة الغربية بالمملكة العربية السعودية بواسطة 6 نظم إنزيمية. وقد اظهرت النتائج وجود تمايز جيني واضح داخل وبين عشائر نبات الحنظل. ظهرت عشرة مواقع جينية كمواقع وحيدة المظهر في كل العشائر، في حين اعتبرت سبعة أخرى (ADH-1, MDH-1, β -EST-1, α -EST-1) داخل العشائر (A = 1.21، P = 31.17%، أقل من تلك الخاصة بالأنواع المعتمدة على التلقيح الحشري (P = 34%؛ He = 0.205، A = 2.67). وكان متوسط التباين الجيني الملحوظ (0.757) أعلى من متوسط التباين الجيني المتوقع (0.415) مما دل على وجود انتقاء طبيعي ضد التماثل الجيني. وكانت قيم معامل تزاوج الأقارب (F) للمواقع الجينية متعددة الأشكال سلبية مما يشير إلى وجود زيادة كبيرة في التوجه نحو التباين الجيني داخل عشائر الحنظل. وقد أكد مخطط الشكل الشجري للقرابة المعتمد على 84 فردا من عشائر الحنظل ذلك التباين السابق ذكره داخل العشائر. وعلى الجانب الآخر سجل التمايز بين العشائر 16.2% من مجموع التباين الكلي، وهذه القيمة المنخفضة مع قيم المسافات الوراثية العالية (D) أظهرت أن التمايز بين العشائر كان منخفضا على العكس من التباين داخلها. وبالإضافة لكل ما سبق ومع تقدير تدفق منخفض للجينات في جميع العشائر باستثناء تلك المجمعه من مدينة مكة، نوصى بصلاحية نباتات الحنظل الموجودة بمكة- من خلال نتائجنا- لبرامج التربية وحفظ الأصول الوراثية.

وتخلص الدراسة إلى أهمية البيئة الصخرية القاحلة في سيناء كمصدر متميز للأكتينوبكتريا ذات قدرات واعدة لإنتاج المضادات الحيوية.